

1 Environmental impacts by fragments released from  
2 nano-enabled products: A multi-assay, multi-  
3 material exploration by “the SUN approach”.

4 *Monica J.B. Amorim, Sijie Lin, Karsten Schlich, José M. Navas, Andrea Brunelli, Nicole*  
5 *Neubauer, Klaus Vilsmeier, Anna L. Costa, Andreas Gondikas, Tian Xia, Liliana Galbis, Elena*  
6 *Badetti, Antonio Marcomini, Danail Hristozov, Frank von der Kammer, Kerstin Hund-Rinke,*  
7 *Janeck J. Scott-Fordsmand, André Nel, Wendel Wohlleben\**

8

9 AUTHOR ADDRESS

10 Mónica J.B. Amorim, University of Aveiro, Department of Biology and CESAM, 3810-193  
11 Aveiro, Portugal (mjamorim@ua.pt)

12 Sijie Lin, College of Environmental Science and Engineering, State Key Laboratory of Pollution  
13 Control and Resource Reuse, Tongji University, Shanghai 200092, China; Center for  
14 Environmental Implications of Nanotechnology, California NanoSystems Institute, University  
15 of California Los Angeles, CA 90095, United States States (lin.sijie@tongji.edu.cn)

16 Karsten Schlich, Kerstin Hund-Rinke, Fraunhofer Institute for Molecular Biology and Applied  
17 Ecology, Dept. Ecotoxicology, Auf dem Aberg 1, 57392 Schmallenberg, Germany.  
18 (Kerstin.Hund-Rinke@ime.fraunhofer.de)

19 José M Navas, Liliana Galbis. INIA, Dept. of Environment, Ctra. De la Coruña Km 7.5 E-28040  
20 Madrid

21 Andrea Brunelli, Elena Badetti, Antonio Marcomini, Danail Hristozov, Dept. of Environmental  
22 Sciences, Informatics and Statistics (DAIS), University Ca' Foscari of Venice, Via Torino 155,  
23 30170 Venice Mestre, Italy (andrea.brunelli@unive.it), (elena.badetti@unive.it),  
24 (marcom@unive.it), (danail.hristozov@unive.it)

25 Nicole Neubauer, Klaus Vilsmeier, Wendel Wohlleben, Dept. Material Physics, BASF SE, 67056  
26 Ludwigshafen, Germany. (wendel.wohlleben@basf.com)

27 Anna L Costa, ISTECCNR, Via Granarolo, 64, I-48018, Faenza, Italy (anna.costa@istec.cnr.it)

28 Andreas Gondikas, Frank von der Kammer, Dept. of Environmental Geosciences, University of  
29 Vienna, 1090 Vienna, Austria (frank.von.der.kammer@univie.ac.at)

30 Andreas Gondikas, current address: Dept. of Marine Sciences, University of Gothenburg, 412-96  
31 Gothenburg, Sweden (andreas.gondikas@gu.se)

32 Janeck J. Scott-Fordsmand, Aarhus University, Dept Bioscience, Vejlsovej 25, PO Box 314, 8600  
33 Silkeborg, Denmark (jsf@bios.au.dk )

34 Tian Xia, André Nel, Division of NanoMedicine, Department of Medicine, Center for  
35 Environmental Implications of Nanotechnology, California NanoSystems Institute, University  
36 of California Los Angeles, CA 90095, United States (txia@ucla.edu; anel@mednet.ucla.edu)

37 Wendel Wohlleben, Dept. Experimental Toxicology and Ecology, BASF SE, 67056 Ludwigshafen  
38

39 KEYWORDS: nanomaterials; nano-enabled products; lifecycle; microplastics; environmental  
40 effects; pigments; carbon nanotubes; polyolefin; cement; epoxy

41

## 42 ABSTRACT

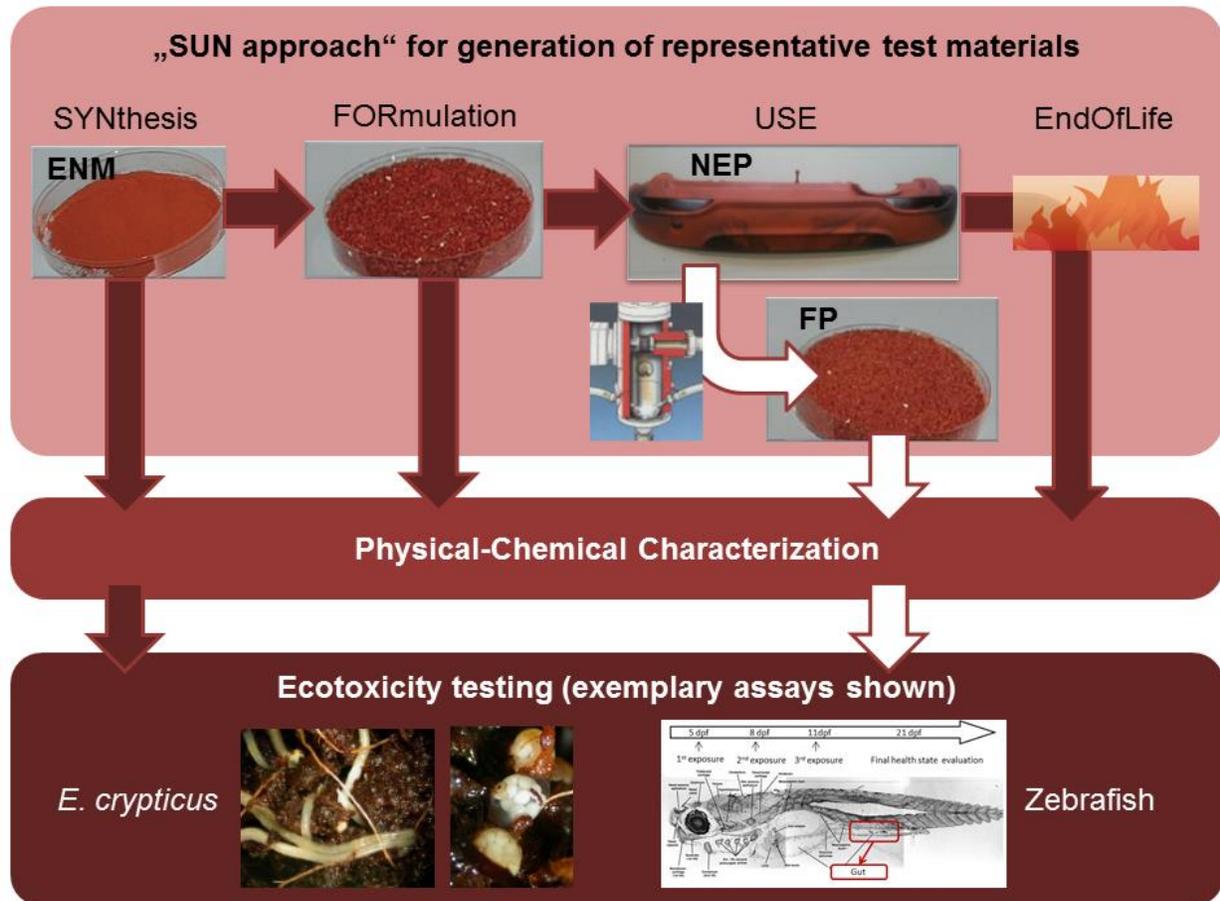
43 Nano-enabled products (NEPs) have numerous outdoor uses in construction, transportation or  
44 consumer scenarios, and there is evidence that their fragments are released in the environment at  
45 low rates. We hypothesized that the lower surface availability of NEPs fragment reduced their  
46 environmental effects with respect to pristine nanomaterials. This hypothesis was explored by  
47 testing fragments generated by intentional micronisation (“the SUN approach”, Nowack et al,  
48 ES&T **50** (2016) 2747). The NEPs were composed of four matrices (epoxy, polyolefin,  
49 polyoxymethylene, cement) with up to 5% content of three nanomaterials (carbon nanotubes, iron  
50 oxide, organic pigment). Regardless the type of nanomaterial and/or matrix used, it was observed  
51 that nanomaterials were only partially exposed at the NEP fragment surface, indicating that mostly  
52 the intrinsic and extrinsic properties of the matrix drove the NEP fragment toxicity.  
53 Ecotoxicity in multiple assays was done covering relevant media from terrestrial to aquatic,  
54 including sewage treatment plant (biological activity), soil worms (*Enchytraeus crypticus*), and  
55 fish (zebrafish embryo and larvae, trout cell lines). We designed the studies to explore the possible  
56 modulation of ecotoxicity by nanomaterial additives in plastics/polymer/cement, finding none. The  
57 results support NEPs grouping by the *matrix* material regarding ecotoxicological effect during the  
58 use phase. Further, control results on nanomaterial-free polymer fragments representing  
59 microplastic had no significant adverse effects up to the highest concentration tested.

## 60 **Introduction**

61 Nanocomposites are a dominant class of nano-enabled products (NEPs), and especially those with  
62 a “durable” matrix, such as rubber, cements, plastics and coatings, are often in outdoor use.<sup>1, 2</sup>  
63 Processes such as wear, tear, manufacturing and shredding (mechanical processes) and photolysis,  
64 hydrolysis, aggressive use environments, thermal decomposition (chemical processes) contribute  
65 to the degradation of nanocomposites<sup>3, 4</sup> and induce releases into the environment at various time  
66 scales along the lifecycle.<sup>5, 6</sup> Pristine Engineered Nanomaterials (ENM) can induce adverse effects  
67 on environmental species in aquatic, sediment and soil compartments.<sup>7</sup> This is relevant for  
68 emission scenarios such as spills during the ENM synthesis phase of the lifecycle, but not for the  
69 NEP use phase. For NEPs made of chemically resilient nanocomposite materials, the only relevant  
70 case for NEP outdoor use, nanocomposite fragments of the NEPs are the dominant physical-  
71 chemical form of environmental releases during the use phase.<sup>8-10</sup> However, to the best of our  
72 knowledge no study has ever reported on environmental effects by NEP fragments. This is a  
73 serious gap in environmental risk assessment of nanomaterials. NEP fragments differ from pristine  
74 ENM in key physical-chemical properties such as composition, size, surface chemistry.<sup>11</sup> Their  
75 environmental release rates on the order of g/year<sup>12</sup> are too low to measure their properties and  
76 environmental effects directly for degrading NEPs, which in a controlled experiment requires  
77 exposure of various organisms up to high doses. To meet the gap, Nowack et al. proposed “the  
78 SUN approach”: NEPs were micronized to generate fragmented products (FP) in kg quantities for  
79 characterization of properties and effects.<sup>13</sup> Our hypothesis for this study, based on limited reports  
80 on mammalian studies, is that FP will have lower environmental impacts compared to pristine  
81 ENMs due to the lower surface availability of ENM contained in FPs.

82 Here we deliver an exploration of ecological effects of multiple NEP-FPs by multiple ecotoxicity  
83 assays, supported by an extensive physical-chemical characterization of the properties of the FPs  
84 generated by “the SUN approach” (**Figure 1**). Specifically, we compare different ENM in the same  
85 matrix, and additionally explore one ENM in different matrices. We selected a balanced choice of  
86 NEPs: this includes highly studied nanomaterials, of which we chose a family of NEPs with  
87 multiple walled carbon nanotubes (CNTs) in polyolefins for conductive functionality / in cement  
88 for electromagnetic shielding functionality / in epoxy for lightweight functionality in automotive  
89 / construction / airplane applications, respectively.<sup>14</sup> Additionally, we selected a family of NEP  
90 made from less well studied ENM from commercially well-established particulate materials,  
91 which are now identified as nanomaterials in regulatory terms, specifically transparent halogen-  
92 organic and inorganic pigments for automotive parts and coatings.<sup>2</sup> These constitute the large  
93 volume reports in the French nanomaterial registry with above 100 ton/y nanoform production or  
94 import in France, far above CNT.<sup>15</sup> Due to the history of widespread use of nanoforms of these  
95 materials in coating, ink, plastic components of consumer products, a historical and ongoing  
96 dispersive emission of NEP fragments into the environment has to be anticipated. We previously  
97 verified that the ENMs are present in and on the FPs, and that even after extensive (365 MJ/m<sup>2</sup>)  
98 UV radiation less than 6 ppb of the ENM content migrates or leaches from the weathered FPs  
99 (WFPs),<sup>16</sup> such that testing of environmental effects can be approximated by testing FP only. This  
100 reasoning is specific to environmentally persistent matrices such as polyolefins or cements, as  
101 tested here. Our study is the first to enable their environmental risk assessment with full  
102 consideration of the nanostructures induced by the ENM content.

103



104  
 105 **Figure 1.** How can we assess environmental impacts from nano-enabled products?  
 106 Representative test materials across all lifecycle phases are needed: The pristine ENM powder  
 107 (here an organic pigment), represents spills during nanomaterial synthesis (SYN), and the pigment-  
 108 in-polymer masterbatch granules represent spills during formulation (FOR). But the nano-enabled  
 109 product (NEP), a 1.4m-wide car bumper in one of our case studies, releases fragments only at very  
 110 slow rates.<sup>12</sup> Specifically to the “SUN approach”, highlighted by white arrows, mechanical  
 111 micronisation generates fragmented products (FP) to represent the real-world NEP releases with  
 112 greatly increased surface and surface-accessible ENM.<sup>13</sup> Photographs illustrate test materials in 10  
 113 cm petri dishes (no ashes remain after incineration in the specific case). Materials representing all  
 114 lifecycle phases were compared regarding their physical-chemical properties. The most relevant

115 materials were compared regarding their ecotoxicity by multiple assays, of which the lower panels  
116 show: Enchytraeidss (soil): *Enchytraeus crypticus*. in culture soil and a detail of the cocoons with  
117 eggs inside, and the Zebrafish (aquatic): *Danio rerio* development assay using water-  
118 accommodated fractions. We assessed six NEPs and their ENM-free control matrices in a  
119 systematic variation of ENMs and of matrices (see Figure 3).

120

## 121 **Materials and Methods**

122 *ENM, synthesis phase and NEP, formulation phase:* The test panel consisted of three ENM and  
123 four matrices. The same nanomaterials have been used previously for extensive human toxicity  
124 testing.<sup>17, 18</sup> “CNT” designates multi-wall CNT, CAS 308068-56-6, and is the identical grade as  
125 the batch distributed as NM400 in the OECD sponsorship program. “OrgPig” designates di-keto-  
126 pyrrolo-pyrrole, CAS 84632-65-5. “Fe<sub>2</sub>O<sub>3</sub>” designates hematite, CAS 1309-37-1 (**Table SI\_1**).  
127 All ENM were formulated into polyolefins (high density polyethylene, HDPE, and polypropylene,  
128 PP). Additionally to PP, the CNT was integrated in three complementary matrices:  
129 Polyoxymethylene (“POM”) nanocomposite identical to the FP used in a human tox study<sup>19</sup>,  
130 cement (“cement”) nanocomposite identical to the FP used in a human tox study<sup>19</sup>, epoxy  
131 (“epoxy”) nanocomposite identical to those applied in the NanoRelease interlab testing (**Table**  
132 **SI\_2**).<sup>20</sup> Comparative testing of environmental weathering releases was performed on the identical  
133 nanocomposites, finding release rates that increase across five orders of magnitude in the order  
134 polyolefin – POM – epoxy – cement.<sup>21</sup>

135 *NEP fragmentation, use phase:* We implemented “the SUN approach” (**Figure 1**) and performed  
136 cryo-milling for all NEPs as listed in **Table SI\_2**. In short, the OrgPig\_PP, Fe<sub>2</sub>O<sub>3</sub>\_PE,

137 CNT\_epoxy, CNT\_PP nanocomposite materials and the PP, PE, epoxy nano-free controls were  
138 frozen to cryogenic temperatures (at -193°C, liquid N<sub>2</sub>) in order to maximize brittleness.<sup>13</sup>  
139 Additionally, in order to remain comparable to earlier human tox investigations on sanding dusts,  
140 limited tests were performed on CNT\_POM and CNT\_cement fragments and their POM, cement  
141 control fragments. These were generated by sanding (1.8 m/s, grit 80, 10 N) of the same NEP batch  
142 that was previously investigated by rat in vivo studies.<sup>19</sup> See SI for fragmentation details.

143 *FP size analysis:* Size distributions have been previously reported.<sup>13</sup> and are included by the  
144 median diameters here. In short, FP were dispersed in a concentration of 1 g/l by sonication in  
145 water containing 0.5 g/l SDS (Sodiumdodecylsulfate), and were characterized by laser diffraction  
146 (Malvern Mastersizer 3000).

147 *FP composition analysis:* The content of ENM in several FP was not known previously, and was  
148 investigated here by ICP-MS for the inorganic ENM and by combustion for the organic pigment.  
149 In the case of CNT, the detection was based on the content of Al and Co in the used CNT which  
150 remains from their production process. The Al and Co content is known from earlier release  
151 investigations on the same batch of CNT\_epoxy.<sup>20</sup>

152 *FP surface chemistry analysis (X-ray photoelectron spectroscopy, XPS + hydrophobicity):*  
153 Surface composition was investigated by XPS to determine element concentrations and content of  
154 ENM at the fragments' surface. Phi XPS 5500 with 300 W monochromatic Al-K alpha radiation,  
155 pass energy for surveys 117 eV. The XPS penetration depth of 10 nm is around three orders of  
156 magnitude smaller than the FP diameters. Identical analysis was performed on pristine ENM. To  
157 evaluate the hydrophobicity, contact angle measurements were performed by placing a sessile drop

158 of water onto a round pressed plate of ENM or FP respectively with a diameter of 3 cm. Triplicates  
159 had standard deviations between 2° and 6°.

160 *FP “biologically accessible” fraction of ENM:* Biologically accessible ENM are defined as those  
161 that are in contact to the surrounding liquids, and are considered as the fraction that can potentially  
162 elicit an ENM-specific effect. FP were dispersed in 1% HCl and shaken for up to 22h. At each  
163 sampling, the suspension was left to settle for 10 minutes, then sampling an aliquot for ICP-MS to  
164 determine Al, Co, and Fe concentrations. The HCl concentration chosen is enough to keep traces  
165 of these elements in a truly dissolved state, while it is not expected to significantly corrode the  
166 organic matrix. The method was adapted from Schlagenhauf et al.<sup>22</sup>

167 *FP “dispersability” in liquid media:* FP were dispersed by testing both bath sonication (30 min,  
168 power 35W) and probe sonication (15 min, power 100W) into different media, specifically:  
169 distilled water (DW) + sodium dodecyl sulfate (SDS) at 20 mM; DW + Suwannee River Humic  
170 acid (SRHA) at 0.2% w/w; Artificial freshwater (AFW, adhered to OECD 203 protocol<sup>23</sup>) + SRHA  
171 at 0.2% w/w. As expected from polyolefin density, flotation of FPs on the water surface was  
172 observed. The results were confirmed within the concentrations range from 10 mg/l to 10 g/l by  
173 means of Dynamic Light Scattering (DLS, multi-angle Nicomp ZLS Z3000 Particle Sizing  
174 System), Centrifugal Separation Analysis (CSA, LUMiSizer 651, L.U.M. GmbH) and number-  
175 based Transient Resistive Pulse Sensing (TRPS, izon qnano).

#### 176 ***Ecotoxicological testing of fragmented material***

177 Testing aimed to cover from aquatic to terrestrial exposure, using a suite of standard organisms  
178 and associated endpoints as these are among the current mandatory framework. As shown in Figure  
179 1, each phase of the lifecycle may result in environmental emission by spills during SYN and FOR,  
180 or by slow micronisation and degradation during USE or finally by landfilling of remaining solids

181 after incineration (polymers) or after destruction (cements), impacting sewage treatment plants,  
182 soils and other compartments.

### 183 *Sewage sludge treatment plant function*

184 Effects on the biological function of a sewage treatment plant (STP) were investigated based on  
185 the OECD Test Guideline 303A (OECD, 2001). A lab-scale STP (behrotest® Laborkläranlage  
186 KLD 4N, Germany) with a denitrification and nitrification reactor and a secondary clarifier was  
187 used. The pristine Fe<sub>2</sub>O<sub>3</sub> and the fragmented Fe<sub>2</sub>O<sub>3</sub>\_PE were added continuously over 10 days with  
188 the synthetic sewage into the denitrification reactor of the model STP. Effects on the biological  
189 function of the STP were assessed by measurements of the elimination rate of dissolved organic  
190 carbon and the determination of nitrate, nitrite and ammonia in the effluent of the sewage treatment  
191 plant, representing the denitrification and nitrification processes. Treatments included from 0.04  
192 mg/L (environmental relevant) to 1.0 mg/L (worst case scenario).

### 193 *Microbial function - Short and long-term effects on the soil microflora*

194 The nitrite content was determined using the short-term potential ammonium oxidation test as  
195 recommended by ISO Guideline 15685 (2012). The effect on ammonia oxidizing bacteria was  
196 determined 24 h and 28 days after application of the nanomaterials (10 and 1000 mg/kg dry matter  
197 soil) into reference soil.

### 198 *Long term effects via sewage sludge spread into agricultural soil*

199 After 10 days continuous addition of the nanomaterial into the STP, the sludge was dewatered and  
200 added to the reference soil in accordance with the German sewage sludge ordinance, which states  
201 5 tons per hectare over 3 years can be spread on agricultural areas. 1.67 g of dry matter sludge  
202 were introduced into the soil, under the assumption of a soil depth of 20 cm and soil density of 1.5  
203 g/m<sup>3</sup>, corresponding to test concentrations of 0.4 and 10 mg/kg dry matter soil. The soil was

204 incubated at 20°C in an incubation chamber in the dark. The long term effect of the nanomaterials  
205 added via sewage sludge on the ammonia oxidizing bacteria was investigated after 30, 60, 100 and  
206 140 days of incubation.

207 *Enchytraeid Reproduction Test (ERT):*

208 Cultures of test species *E. crypticus* (Oligochaeta: Enchytraeidae, **Figure 1**) were kept in agar  
209 plates for several years.<sup>24</sup> The standard LUFA 2.2 natural soil (Speyer, Germany) was used.  
210 Spiking followed the recommendations for nanomaterials<sup>25,26</sup> The standard guideline (ISO 2004;  
211 OECD 2004) was followed, but instead of using adults, 10 synchronized age (17-18 days)  
212 organisms were used. Exposure concentrations ranged from 0-3200 mg/kg soil dry weight for both  
213 the ENM and the FP, i.e. for the FP exposure there was a corresponding lower % of ENM. Test  
214 ran at 20 °C and 16:8h photoperiod during a period of 28 days. Endpoints assessed included  
215 survival (number of adults) and reproduction (number of juveniles).

216 *Hatching, survival and growth test on zebrafish embryos and larvae*

217 Zebrafish embryos and larvae were used to assess the hazard potential in a model organism  
218 frequently used in nanomaterial safety studies (**Figure 1**). Motivated by FP hydrophobicity and  
219 lack of dispersability, sample preparation for aquatic compartment testing (see details in SI)  
220 targeted the water-accommodated fraction as recommended by an OECD workshop.<sup>27</sup> Healthy  
221 embryos at the same developmental stage (2 hour post fertilization, hpf) were selected and placed  
222 in 96-well plates with one embryo per well.<sup>28,29</sup> One hundred microliters of CNT, or FP of CNT-  
223 cement and CNT-POM was added to the wells at 4 hpf. To achieve robust statistical calculation,  
224 five replicate trials were carried out, each using 12 embryos. Observations of adverse biological  
225 outcomes, including hatching interference, phenotypic abnormalities and mortality (necrosis of the  
226 embryos), were carried out every 24 h for five consecutive days.

227 Using our previously developed pulse exposure procedure,<sup>30</sup> we further investigated the hazard  
228 potential of the CNT and FP from CNT-nanocomposite in zebrafish larvae. As shown in **Figure**  
229 **SI\_5**, the pulse exposure was conducted using groups of 30 growth-synchronized larvae at 5, 8  
230 and 11 days post fertilization (dpf). The exposure was performed in Petri dishes for 6 h each time  
231 by immersing the larvae in 3 mL of the suspension of CNT or FP. The larvae were maintained in  
232 standard aquarium tanks for further observation of development and survival until 21 dpf. The  
233 overall health status of the larvae was assessed at 14 dpf based on gross morphology, body length  
234 and weight, and number of calcified vertebrae. To assess morphology, larvae were anesthetized in  
235 0.02% tricaine and embedded in low-melt agarose gel. This allowed positioning to obtain lateral  
236 views to assess morphology, body length, and fin structure.

#### 237 *Cytotoxicity on fish cell lines*

238 Two fish derived cell lines were used: RTL-W1<sup>31</sup> and CLC.<sup>32, 33</sup> For details see SI. Cells were  
239 exposed for 24 h to appropriate concentrations of the original NM or of the FP (0.78 to 100 µg/mL)  
240 using 96 well plates. Two independent experiments were carried out, with treatments applied in  
241 triplicate in each experiment, Fresh suspensions were prepared for each experiment in milliQ water  
242 at 10 mg/ml by sonication for 20 min in an ice-water bath using a probe sonicator (Vibra cell  
243 VCX130, Sonics & Materials Inc., Newtown, CT, USA) (2 mm microtip, 80% of amplitude  
244 continuous mode). 75 µl were then mixed with 25 µl of bovine serum albumin (BSA, 80 mg/l)  
245 solution, added to 7.4 ml of culture medium, and sonicated for 10 min in a bath sonicator (S 40 H  
246 Elmasonic, Elma, Germany). Cells were exposed to ½ serial dilutions of this suspension in culture  
247 medium. Cytotoxicity was measured by three different assays.<sup>34, 35</sup>

248

## 249 **Results & Discussion**

250 Enabled by the “SUN approach” (**Figure 1**), we followed nanomaterials along their lifecycle, both  
251 by physical-chemical and by ecological methods. The pristine ENM, representing the SYN (as  
252 SYNthesized) phase, and the fragmented product, representing releases during the USE (in USE)  
253 phase, were characterized in terms of size, composition, and surface properties (**Figure 2** and  
254 **Table SI\_3**). Additionally, selected properties were available from our previous publications or  
255 from gap-filling original data for the FOR (FORMulation) and EOL (End Of Life) phases. **Figure**  
256 **3** thus constitutes a “lifelog” of nanomaterials from cradle to grave.

257 Our specific understanding of lifecycle phases differentiates:

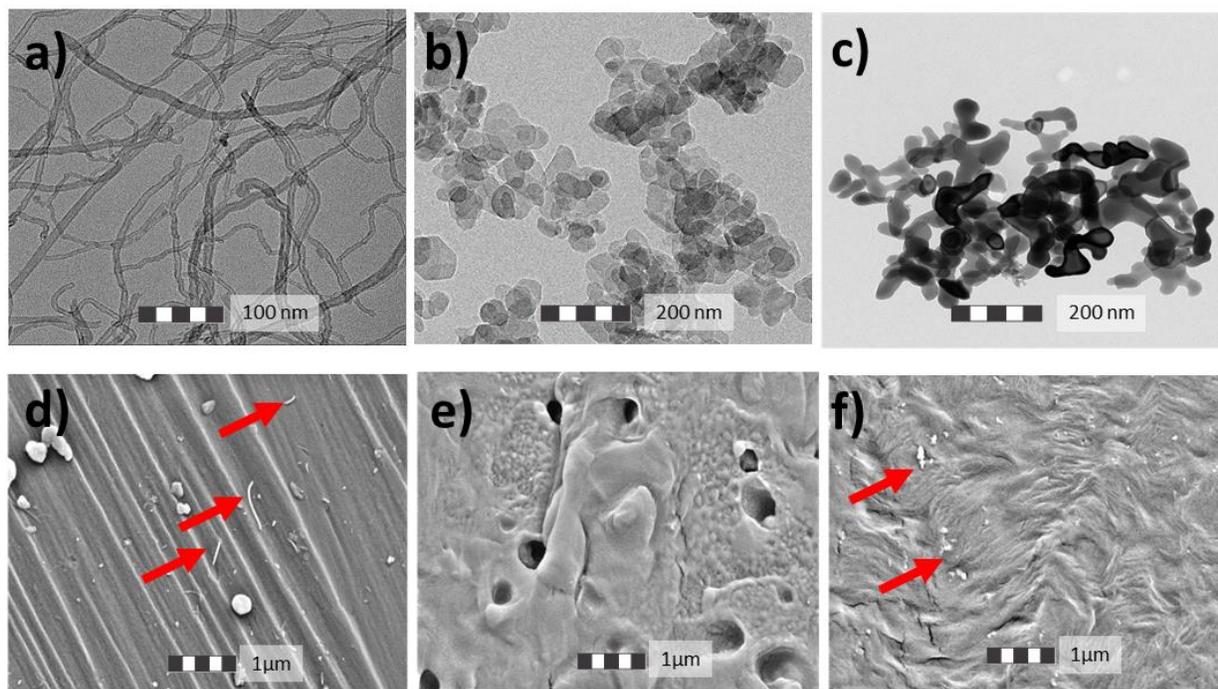
- 258 • SYN = synthesis of the pristine ENM, typically as powder. Industrial process by chemical  
259 operations. Risk assessment prioritizes occupational human aerosol exposure and  
260 environmental emissions by spills and waste streams.
- 261 • FOR = formulation of the ENM as an additive in a product matrix. Industrial manufacturing  
262 process that produces a nano-enabled product (NEP). Our case studies used ENM  
263 masterbatch (for thermoplastic polymers PP, PE, POM), or ENM suspensions in liquid  
264 precursors (epoxy polymer and cement).
- 265 • USE = consumer use of the NEP. Risk assessment needs to consider various product-  
266 specific scenarios with durations between days and decades. For our specific case studies,  
267 environmental emissions of FP by weathering, wear and tear during outdoor use take  
268 priority.

- 269 • EOL = end of life of the NEP by recycling or disposal by incineration or landfill. For our  
270 specific case studies Table 3 reports the results measured for incineration of epoxy<sup>36</sup> and  
271 PP<sup>37, 38</sup> or landfill of cement FP.

272 Previously we reported the FP size as a basic descriptor.<sup>13</sup> Here we go further to characterize the  
273 composition and surface chemistry by several descriptors that are most relevant for nanoscale- or  
274 micro-particles. The actual ENM content in FP (USE phase) was found to be minimally above the  
275 ENM content of the FOR phase, resulting in a small kink in the “nanomaterial lifelog” between  
276 FOR and USE phase (**Figure 3**, panel “composition”). The increase in ENM content correlates  
277 roughly with the brittleness of the matrix, as it is least with PP and PE, higher with epoxy and  
278 cement. Transmission Electron Microscopy (TEM) scans of iron oxide pigments extracted from  
279 Fe<sub>2</sub>O<sub>3</sub>\_PE fragments showed no significant change of nanomaterial structure due to the  
280 fragmentation process (ultramicrotomy data not shown). As the changes of composition are  
281 minimal between FOR phase and USE phase (**Figure 3** and **Table SI\_3**), whereas only the USE  
282 phase potentially releases fragmented product by diffuse emissions into the environment, we  
283 focused testing on the SYN and USE phases. Sizes after sieving were above the inhalable range  
284 for all cryo-milled FP (**Figure 3**, panel “size”), in accord between laser diffraction, suspension-  
285 based analytics and microscopy.<sup>13</sup> The amounts of the sieved fragments were below 0.5% of  
286 cryomilling output, and were provided for ecotoxicological testing. Additionally, FPs produced by  
287 sanding CNT\_POM\_FP were consistent in size with the other FP generated by cryo-milling, as  
288 the CNT\_POM\_FP had < 1 % passed through a 2.7µm filter, and < 60 ppm were below 0.1 µm  
289 (both w/ and w/o CNT); 3 % of the CNT\_cement\_FP passed through a 2.7 µm filter, and 100 ppm  
290 were below 0.1 µm.<sup>19</sup> High resolution images reveal protrusions of the inorganic Fe<sub>2</sub>O<sub>3</sub> pigment  
291 and CNTs at the surface of corresponding fragments (red arrows in **Figure 2**), whereas the lower

292 concentration and lower electron density of OrgPig caution against an interpretation of absence of  
293 protrusions from SEM results.

294



295 **Figure 2** SEM scans to characterize surface structures of a)-c) ENM (representative for lifecycle  
296 SYN phase) and d)-f) fragments of NEP containing the same ENM (FP, representative for lifecycle  
297 USE phase). a) CNT; b) halogen-organic diketopyrrolopyrrole pigment red 254 (OrgPig); c) Fe<sub>2</sub>O<sub>3</sub>  
298 pigment red 101; d) FP of nanocomposite CNT\_epoxy; e) FP of nanocomposite OrgPig\_PP; f) FP  
299 of nanocomposite Fe<sub>2</sub>O<sub>3</sub>\_PE. Red arrows highlight protrusions of ENM on FP surfaces. See  
300 **Figure SI\_1** for pure matrix control fragments and whole-particle scans.

301

302 “Protrusions”<sup>22, 39</sup> of ENM on the surface of fragments were investigated by multiple approaches:  
303 surface chemistry (XPS, elements and C(1s) line shift identification), hydrophobicity (sessile water  
304 drop contact angle), morphology Scanning Electron Microscopy (SEM). As hypothesized in the

305 introduction, the surface of all investigated FP materials is mainly composed of carbon (**Table**  
306 **SI\_3** and **Figure 3**, panel “surface chemistry”). In case of the epoxy-based materials, further  
307 elements such as carbon and nitrogen are present in higher contents as well. The XPS  
308 quantification of different species by C(1s) line shift (**Figure SI\_2**) analysis finds a *surface* content  
309 of CNT in CNT\_PP\_FP of 4.3%, which is slightly increased above the *bulk* content determined by  
310 ICP-MS of 2.7%. Also the Al and Co acid leaching analysis showed that the CNT are accessible  
311 to the liquid phase on the surface of the CNT\_PP fragments, as these released Co and Al (**Figure**  
312 **SI\_3**). The biologically accessible concentration of Al was deduced to be 27.0  $\mu\text{g}/\text{m}^2$  on FP with  
313 CNT and 7.6  $\mu\text{g}/\text{m}^2$  without; the respective values for Co were 2.3  $\mu\text{g}/\text{m}^2$  and 0.1  $\mu\text{g}/\text{m}^2$  (**Table**  
314 **SI\_4** and calculation there). Correcting for the control background, and using the diameter, density  
315 and metal content in this specific CNT grade, we deduce from the acid leaching that 6.2 % of the  
316 surface area provide contact to CNT. Considering the approximations made, this value is in  
317 excellent accord with the XPS result and predicts that protrusions of ENM modulate the surface  
318 chemistry of FP, but do not dominate it. For CNT in epoxy and for  $\text{Fe}_2\text{O}_3$  in PP however, leaching  
319 results are convoluted by contaminations of the matrix with the target analytes (Al, Fe) (results in  
320 the SI). Further work will aim at determining whether these metals are embedded in the NEP or if  
321 the NEP fragmentation process contaminates the FP. The same methods of XPS line shift, tracer  
322 ion release and TEM analysis, found significant protrusions on brittle CNT\_epoxy<sup>22</sup> or  
323 CNT\_cement,<sup>39</sup> but low prevalence in tough materials such as CNT\_POM or CNT\_PP.<sup>39, 40</sup> This  
324 is in accord with our results.

325 For organic pigment, XPS line shape analysis identifies an enrichment at the FP surface with up  
326 to 9% on the FP surface compared to 0.21% within the FP bulk (**Figure SI\_2**), but may be less  
327 reliable because only CNTs emit C(1s) photoelectrons at a distinct energy shift, whereas the

328 organic pigment signal overlaps with the PP matrix signal. The water contact angle was considered  
329 as simple measure of surface-mediated (extrinsic) interactions. The contact angle results highlight  
330 that the FP surface hydrophobicity is clearly dominated by the matrix, not by the ENM (**Figure 3**  
331 panel “surface chemistry” and **Figure SI\_7**).

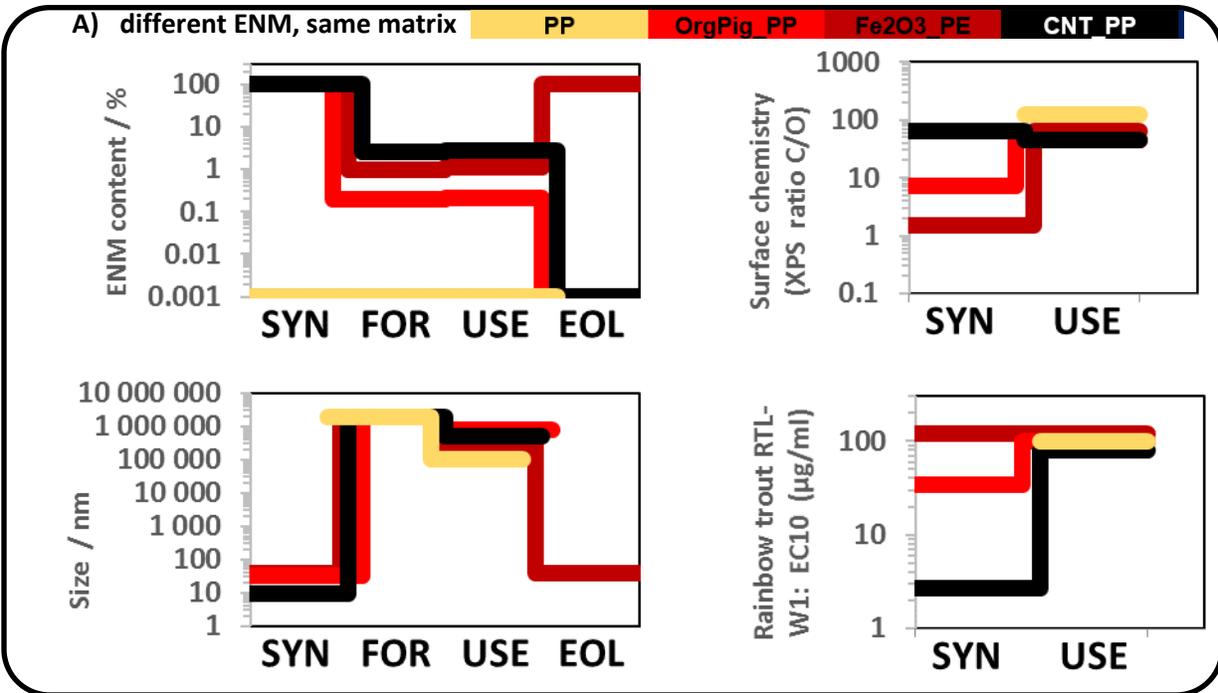
332 Surface chemistry controls nano-bio-interactions, but also impacts fate and transport. In  
333 biologically irrelevant media such as a 1:1 acetone:water mixture, the FP can be solubilized and  
334 dispersed, but in environmentally relevant media such as artificial fresh water with or without  
335 natural organic matter the FP remain agglomerated and tend to float (especially the polyolefins PE  
336 and PP). In accordance to the results from laser diffraction reported earlier,<sup>13</sup> no significant fraction  
337 below 1µm diameter was detected by measurements with the nanospecific methods of DLS and  
338 CSA, using either SDS or SRHA as dispersants. By the nanospecific counting method of TRPS,  
339 all FP showed the presence of fragments in a size range between 200 and 800 nm, but the number-  
340 based concentrations ranged between  $10^6$  and  $10^7$  #/ml for 10g/l FP suspensions (**Table SI\_5**).  
341 This converts to a low ppm level mass content of fragments with diameters below 1µm.

342 In a very condensed presentation of size, surface chemistry and composition, **Figure 3** summarizes  
343 the dramatic changes of physical-chemical properties along the NEP lifecycle.

344

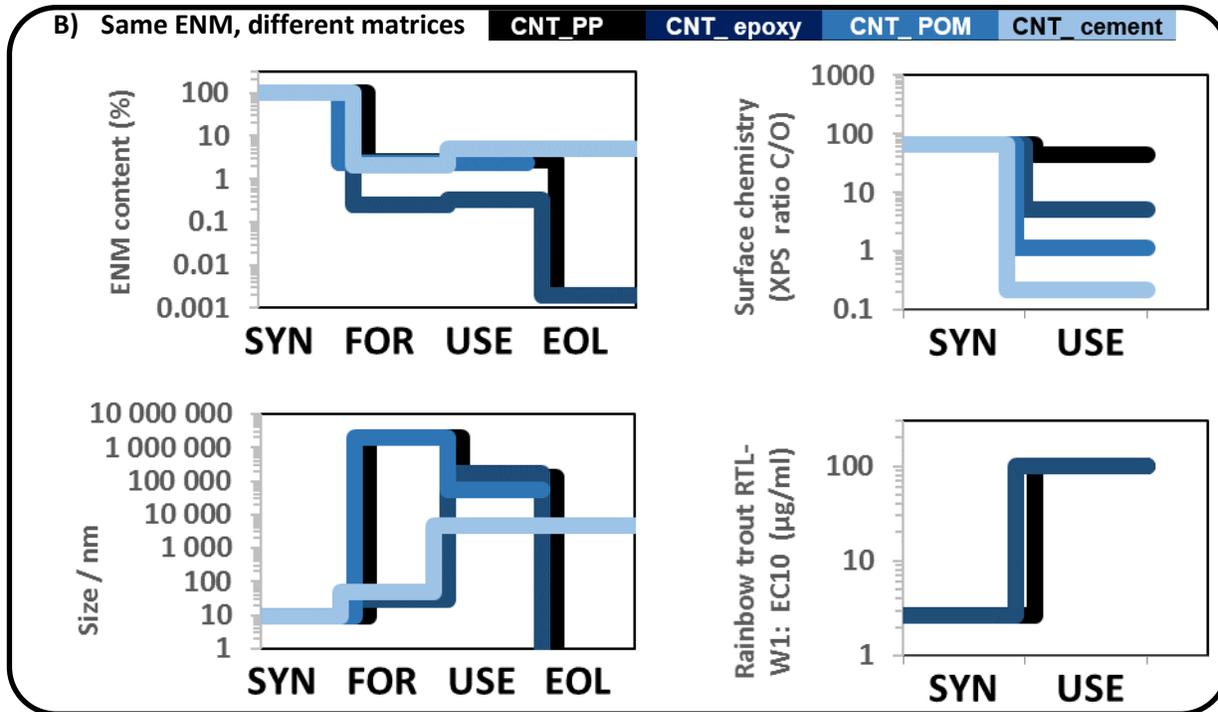
345

346



347

348



349

350

351

352

**Figure 3** Tracking nano-enabled products along their lifecycle stages SYNthesis, FORMulation, USE, End-Of-Life: A) different ENM, same matrix; B) same ENM, different matrices. Panels summarize the key descriptors of Size (median diameter), Composition (ENM content), Surface

353 Chemistry (C/O ratio) and Ecotoxicity (represented by our most sensitive assay, rainbow trout  
354 RTL-W1). Ecotoxicity EC10 values are lower limits for the USE phase. Color code of NEPs is  
355 indicated above the figure and matches Table 1. See beginning of results section for the scenario  
356 assumptions on SYN-FOR-USE-EOL phases.

357

358

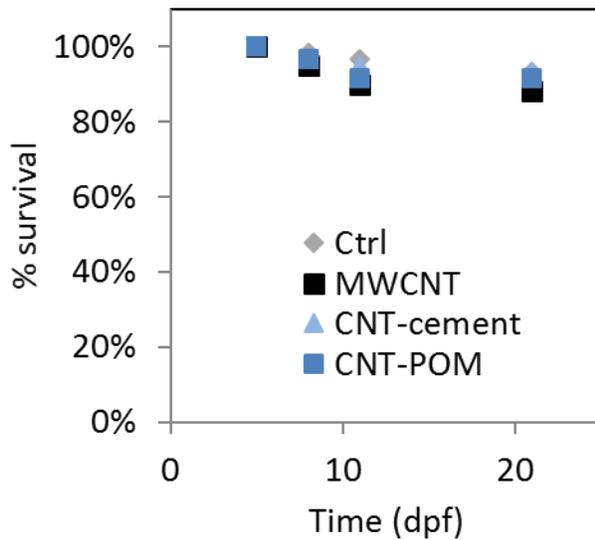
### 359 *Ecotoxicity effect characterisation*

360 No hatching or survival rate of zebrafish *embryos* were affected by CNT or the FP (**Figure SI\_5**),  
361 likely due to low heavy metal content from the CNT and little to no penetration through the chorion  
362 of the embryos. With the pore size of the chorion being at the range of 200~500 nm, it was  
363 anticipated that there was little to none penetration of CNT and FP into the chorionic space. As  
364 shown in **Figure 4**, both CNT and CNT-composite showed little to no toxicity as reflected by the  
365 survival and growth of zebrafish *larvae* subject to aqueous exposure. Even though in the zebrafish  
366 assay CNT were protruding and entirely biologically accessible after shaking in solution or  
367 sonication from CNT-cement\_FP (**Figure SI\_4**), whereas there was no protrusion from CNT-  
368 POM\_FP, the composites showed no adverse effects on zebrafish embryo hatching as well as  
369 survival and growth of zebrafish larvae. These results show that CNT nanocomposites do not  
370 appear to have negative impact on zebrafish. Zebrafish embryo hatching was previously  
371 demonstrated to be affected by heavy metal ions, such as  $Zn^{2+}$ ,  $Cu^{2+}$ ,  $Ni^{2+}$  and  $Cr^{3+}$ .<sup>28</sup> The lack of  
372 hatching interference observed here suggests that the heavy metal leaching, observed only in HCl  
373 to reach ppb levels during 22h (**Figure SI\_3**), remained low during the test but may contribute to  
374 long-term effects in the environment. One may differentiate that only free ENM are biologically

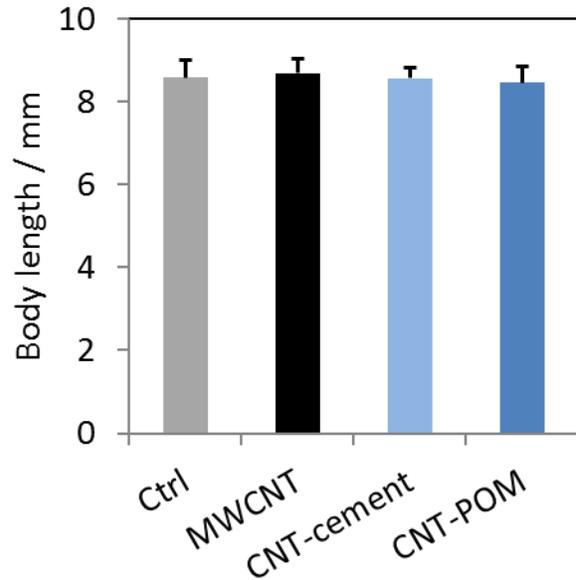
375 available in the sense that they can be taken up, whereas ENM protrusion on a larger fragment are  
376 only accessible in the sense that they interact with e.g. the chorion or cell membrane.

377

378 **A)**



**B)**



379

380 **Figure 4.** Aquatic compartment screening on zebrafish *larvae*: Both A) survival (standard  
381 deviation of N=30 replicates were determined for each time point and were below 5% for all) and  
382 B) body length at day 14 showed no statistically significant differences between the control and  
383 exposure of CNT and FP. Pulse exposure was performed on 5, 8 and 11 dpf (day post fertilization)  
384 (see scheme in **Figure 1**). See **Figure SI\_5** for zebrafish *embryo* development and growth results.

385

386

387 Results of the toxicity of the original material and of the FP (SYN and USE phases, respectively)  
388 on RTL-W1 cells, represented as the most sensitive assay, are summarized by **Figure 3** and **Figure**  
389 **SI\_6**. In both fish cell lines, RTL-W1 and CLC, the original NM (SYN phase) caused decreased  
390 cell viability. The lowest EC50 was detected in RTL-W1 cells by means of the 5-

391 carboxyfluorescein diacetate acetoxymethyl ester (CFDA-AM) assay,  $23.56 \pm 5.0$   $\mu\text{g/ml}$ . The  
392 toxicity in the CLC cells was below EC50 hence this was not calculated. The exposure of cells to  
393 the FP (USE phase) did not cause any toxicity, even at the maximal concentration used, 100  $\mu\text{g}$   
394 FP/ml. However, 100  $\mu\text{g}$  FP/ml contains nanomaterials at  $<5$   $\mu\text{g/ml}$ . In addition to this low  
395 concentration of NM, the low density of FPs causes their flotation and hence the contact with the  
396 cells was minimal. Other alternatives, as cells in suspension instead of attached to the bottom of  
397 the plates or an inverted culture should be tested in future to enhance the sensitivity of the assay,  
398 as demonstrated for PP-based incineration-FPs.<sup>41, 42</sup>

399 On ammonia oxidizing bacteria, the reference products and the pristine nanomaterials Fe<sub>2</sub>O<sub>3</sub> Red  
400 101, Organic Pigment Red 254 and CNT showed no effects at test initiation and after 28 days. A  
401 comparable result was found for the fragmented products Fe<sub>2</sub>O<sub>3</sub>\_PE, OrgPig\_PP, and CNT\_epoxy,  
402 although for Fe<sub>2</sub>O<sub>3</sub>\_PE and CNT\_epoxy at 1000 mg/kg dry matter soil inhibitory effects of 23%  
403 and 38%, respectively, were determined at test start. However, this effect could not be confirmed  
404 after 28 days of incubation. The results are presented in **Table SI\_4**. Thus, after 28 days, EC50  
405 values were above 1000 mg/kg dry matter soil for all products tested. No impact occurred on the  
406 biological function/microbial activity of the sewage sludge neither on the elimination of dissolved  
407 organic carbon nor on the denitrification and nitrification processes. There were no effects due to  
408 the pristine and fragmented products observed on the activity of the ammonia oxidizing bacteria  
409 over 140 days after application of the nanomaterials via sewage sludge into soil.

410 Results for *E. crypticus* showed no effects on survival for any of the FP (**Figure 3, Figures SI\_7**  
411 **and SI\_8**). Effects in terms of reproduction showed increased sensitivity, with the pure NM (SYN  
412 phase) having different EC10, specifically OrgPig had a measurable effect (EC10=2009mg/kg),  
413 but not Fe<sub>2</sub>O<sub>3</sub> or CNT. All FP, both NM-containing FP and the pure polymer FP (pure PE, pure

414 epoxy, pure PP) caused no effect in the tested range (0-3200mg/kg). Such results are not  
415 unexpected given the low degradability of the tested matrix or the low fraction of NM embedded  
416 in the matrix. The added characterization of the NEP further confirms the hypothesis. Of note, for  
417 OrgPig\_FP, a 10% reduction in reproduction occurs for 800 mg/kg although no effects occurred  
418 at higher concentrations. A non-monotone dose response has been reported before for Ag<sup>26</sup> and  
419 Ni<sup>43</sup>. This could be related to the potentially reduced agglomeration at lower concentrations hence  
420 higher exposure and effects. However, we do not have evidence to support this hypothesis for the  
421 FP with or without embedded ENMs.

422 **Figures 3** and **Table 1** summarize the results of our explorations: At the ENM concentrations  
423 tested, the NEP matrix properties dominated the intrinsic and extrinsic FP properties, including  
424 their ecotoxicity in a wide range of assays covering sludge, soil, and fish. Two orthogonal  
425 comparisons help to identify the mechanisms:

- 426 a. The surface chemistry, hydrophobicity and the size of fragments assimilate in the USE  
427 phase of different ENM in the same matrix. (**Figure 3a**, **Figure SI\_7**)
- 428 b. Conversely, for one type of ENM in NEPs with different matrices, the surface chemistry  
429 and size of USE phase fragments diversifies. (**Figure 3b**)

430

431

432 **Table 1:** Effects of USE phase Fragmented Product (FP) tested in various ecotoxicity assays.<sup>1</sup> No  
 433 FP elicits a significant effect. NT: Not Tested.

	PP	OrgPig_ PP	Fe <sub>2</sub> O <sub>3</sub> _ PE	CNT_ PP	CNT_ epoxy	CNT_ POM	CNT_ cement
RTL-W1 EC10	> 100 µg/mL	> 100 µg/mL	> 100 µg/mL	> 100 µg/mL	> 100 µg/mL	NT	NT
CLC EC10	> 100 µg/mL	> 100 µg/mL	> 100 µg/mL	> 100 µg/mL	> 100 µg/mL	NT	NT
Soil microflora EC10	> 1000 mg/kg	> 1000 mg/kg	> 1000 mg/kg	NT	> 1000 mg/kg	NT	NT
STP biology EC10	NT	NT	> 6000 mg/kg	NT	NT	NT	NT
E. crypticus EC10 repro.	> 3200 mg/kg	>3200 mg/kg	> 3200 mg/kg	> 3200 mg/kg	> 3200 mg/kg	NT	NT
ZF embryo NOAEL	NT	NT	NT	NT	NT	> 5 mg/ embryo	> 5 mg/ embryo
ZF larvae NOAEL	NT	NT	NT	NT	NT	> 50 mg/mL	> 50 mg/ mL

434  
 435

---

<sup>1</sup> RTL-W1, fibroblast from biliary ducts of rainbow trout. CLC, macrophage cell line from carp,. *Enchytraeus crypticus* survival is not affected, results reported are EC10 of reproduction. Soil microflora, short-term potential ammonium oxidation test based on ISO Guideline 15685. STP biology, biological function of a sewage treatment plant (STP) based on OECD Guideline 303A. ZF embryo, zebrafish embryos exposed to the water-accomodated fraction of the FP mass indicated, assessing hatching percentage and survival rate vs. control embryos. ZF larvae, zebrafish larvae exposed repeatedly to the water-accomodated fraction of the FP concentration indicated, assessing body length and survival rate vs. control larvae.

436 Both categories of lifecycle-induced ENM transformation, assimilation of properties and  
437 diversification of properties by product formulation and fragmentation, have been predicted earlier  
438 by Mitrano et al.<sup>10</sup> Here we studied specific examples with polymeric and cementitious matrices  
439 with organic, carbonaceous and inorganic ENM, and explore their implications for environmental  
440 hazards. In both transformation categories, we find that the physical-chemical properties of the  
441 bulk matrix predominantly determine the physical-chemical properties of the fragments released  
442 during USE, with little modulation by the ENM that was embedded in the NEP. With the specific  
443 ENM and matrices, the ecotoxicity of fragments released during the lifecycle of NEPs is not  
444 affected by the ENM.

445 However, the validity of each individual ecotoxicity assay –that were not developed for FP testing–  
446 is challenged by one or other FP property. For some materials (low density of polyolefins)  
447 floatation limits the delivered dose; this is, however, no issue for FP testing of epoxy, POM and  
448 cement, which are denser than water. Further, the size range of the tested fragments is larger than  
449 generally adequate for the tests and species. To reduce this uncertainty, we sieved away 99.5% of  
450 the larger particles so that exposure was targeted to smallest sizes. And yet, for organisms like *E.*  
451 *crypticus*, the FP sample contains fragments that can be larger than the organisms themselves. A  
452 detailed discussion can be found in a recent opinion paper.<sup>44</sup> Enchytraeids and other oligochaetes  
453 are exposed via soil ingestion and skin adsorption, hence in this case risk is reduced or absent via  
454 exposure. Also the absence of adverse effects on the microbial soil compartment are at least  
455 partially due to the reduction of exposure, because the thermoplastic matrices have low  
456 degradation and diffusion.<sup>9, 45</sup> No effects on soil microflora were found for relatively long periods  
457 up to 140 days, but also a high bacterial resilience may contribute to this absence of effects.  
458 Follow-up options include long term aging of FPs before testing or much longer term biota

459 exposure. Additional species can be added, e.g. filter feeding marine organisms may be susceptible  
460 both to direct effects of microplastic particles<sup>46</sup> or to vector effects of adsorbed toxicants.<sup>47</sup> One  
461 may speculate that toxicant affinity is modulated by the surface chemistry, even if we found surface  
462 chemistry to be dominated by the matrix, not by the ENM.

463 There is no directly comparable study on nanocomposites. One paper reported an unconventional  
464 ecotoxicity assay by exposing fruit flies to an environmentally aged CNT-composite, finding no  
465 specific effects.<sup>48</sup> Seven papers assessed the toxicity for humans by sanding fragments released  
466 from NEPs, and focused on inhalation exposure<sup>19, 22, 49-53</sup> and oral exposure<sup>54</sup>. For paints and  
467 plastics NEPs with silica, titania and CNT additives, the studies report no additional toxicity by  
468 the ENM in the primary target organ,<sup>19, 22, 49-54</sup> only one study found secondary effects in the  
469 liver<sup>53</sup>. Models for environmental risk assessment currently assume the fate and effects of NEP to  
470 be determined by ENM properties.<sup>6</sup> This assumption needs to be revisited in light of the present  
471 finding of NEP environmental effects determined by *matrix* properties, and of their significant  
472 differences in key physical-chemical properties such as composition, size, surface chemistry.<sup>11</sup> We  
473 designed the studies to explore the possible modulation of ecotoxicity by nanomaterial additives  
474 in plastics/polymer/cement. We found no such modulation despite a systematic variation of (Fig.  
475 3A) different nanomaterials and (Fig. 3B) different matrices and (Table 1) test organisms  
476 representing different environmental compartments. In a separate paper, the Predicted  
477 Environmental Concentrations (PEC) based on material-flow exposure models for nanomaterials  
478 and Predicted No Effect Concentrations (PNEC) were estimated with species sensitivity  
479 distribution models. PEC/PNEC risk ratios were obtained for the case studies, species and  
480 environmental compartments described above and indicate no risks based on the given data.<sup>55</sup>

481 With regard to our primary motivation by nanomaterial risk assessment, the present results lend  
482 support to grouping of ecotoxicity during the USE phase: the group would be defined by the *matrix*  
483 material, potentially limited to a certain maximum ENM content that is relevant for ENM  
484 application as functional additive (typically below 5%). The ecotoxicity of fragments emitted  
485 during the USE phase would be read across from the pure matrix material. The grouping by matrix  
486 would probably fail for easily hydrolysible matrices,<sup>8</sup> which would release free ENM and were not  
487 tested here, and for NEPs using dissolving ENM such as Ag, CdTe, which were tested and  
488 reviewed elsewhere.<sup>9, 56-58</sup> Further, secondary fragmentation and release of free ENM may occur  
489 during transport to the final sink in soils or sediments over time scales longer than the standard  
490 test duration for organisms. However, secondary ENM release by weathering and migration was  
491 independently tested on the identical NEP, and found strong containment by the polyolefin  
492 matrix.<sup>16</sup> Another experimental study on tire wear found up to 0.045% releases of free ENM for  
493 secondary UV/hydrolysis degradation, as this only affects a sub- $\mu\text{m}$ -thin surface layer of many-  
494  $\mu\text{m}$ -diameter FP particles.<sup>59</sup> Further, any secondary fragmentation by UV degradation is again  
495 mostly determined by the matrix, with benchmarks of labile (epoxy), intermediate (Polyamide,  
496 POM) and resilient (PE, PP, cement) matrices.<sup>21</sup> Thus, grouping by matrix can be applied to a wide  
497 range of matrices and ENM that is delimited by the intended persistence during use.

498 As a secondary interpretation, one may identify the FP material as microplastics. The size range  
499 and composition match the definition of e.g. the Danish EPA.<sup>60</sup> Plastics, including commodity  
500 plastics with widespread use, are routinely colored with pigments such as  $\text{Fe}_2\text{O}_3$ , whereas the  
501 Organic Pigment Red 254 is rather a high-performance pigment class, and CNTs are currently  
502 lower volume material.<sup>15</sup> Our results on PP, POM, epoxy and PE, with and without ENM, can be  
503 interpreted as one of the first studies on the effects of microplastics on organisms in soils, fish, and

504 on the sewage treatment plant biological activity. We found no significant effects up to the highest  
505 concentration tested. Of note, considerations of physical-chemical similarity as proposed by Hüffer  
506 et al.<sup>11</sup> would position our mechanically shredded plastics with hydrophobic, irregular surfaces  
507 and low aquatic dispersability as more similar to real world secondary microplastics of polyolefin  
508 (packaging), polyamide (fishery nets)<sup>61</sup> or rubber (tire wear)<sup>60</sup> than charge-stabilized polystyrene  
509 beads that are simply more convenient in (eco)toxicity assays.<sup>62</sup> “The SUN approach” thus  
510 contributes to environmental hazard screening of both nano- and micro-structures.

511

512

513 **Acknowledgement:** This work was supported by the project on Sustainable Nanotechnologies  
514 (SUN) that receives funding from the European Union Seventh Framework Programme  
515 (FP7/2007-2013) under grant agreement n° 604305.

516

517 **Supporting Information:** additional SEM scans of shape and surface structure of FPs;  
518 composition and preparation of nanomaterials and NEPs; micronisation of NEPs to FPs; physical-  
519 chemical properties of FPs in tabular format expanding the graphical representation in Figure 3;  
520 FP surface chemistry analysis by XPS line shape and acid leaching; FP dispersability in ecological  
521 media; preparation of FP water-accomodated fractions for zebrafish testing; zebrafish larvae  
522 results; soil microorganism results; cytotoxicity results; *E. crypticus* results.

523

524

525 **Glossary / List of abbreviations**

- 526 AFW: Artificial freshwater
- 527 BSA: Bovine serum albumin
- 528 CFDA-MS: 5-carboxyfluorescein diacetate acetoxymethyl ester
- 529 CNTs: Carbon nanotubes
- 530 CSA: Centrifugal separation analysis
- 531 DLS: Dynamic light scattering
- 532 dpf: days post fertilization
- 533 DW: Distilled water
- 534 EOL: End of life
- 535 ENM: Pristine engineered nanomaterials
- 536 ERT: Enchytraeid reproduction test
- 537 FOR: Formulation
- 538 FP: Fragmented products
- 539 HDPE: High density polyethylene
- 540 hpf: Hour post fertilization
- 541 NEPs: Nano-enabled products
- 542 OrgPig: Di-keto-pyrrolo-pyrrole
- 543 PE: polyethylene
- 544 PEC: Predicted environmental concentrations
- 545 PNEC: Predicted no effect concentrations
- 546 POM: Polyoxymethylene
- 547 PP: Polypropylene
- 548 SDS: Sodiumdodecylsulfate
- 549 SEM: Scanning Electron Microscopy
- 550 SRHA: Suwannee river humic acid
- 551 STP: Sewage treatment plant

552 SYN: Synthesis  
553 TEM: Transmission Electron Microscopy  
554 TRPS: Transient resistive pulse sensing  
555 WFPs: Weathered fragmented products  
556 XPS: X-ray photoelectron spectroscopy  
557

558 **References**

- 559 1. Stark, W.; Stoessel, P.; Wohlleben, W.; Hafner, A., Industrial applications of  
560 nanoparticles. *Chemical Society Reviews* **2015**, *44*, (16), 5793-5805.
- 561 2. Wohlleben, W.; Punckt, C.; Aghassi-Hagmann, J.; Siebers, F.; Menzel, F.; Esken, D.;  
562 Drexel, C.-P.; Zoz, H.; Benz, H. U.; Weier, A.; Hitzler, M.; Schäfer, A. I.; Cola, L. D.;  
563 Prasetyanto, E. A., Nanoenabled Products: Categories, Manufacture, and Applications: Protocols  
564 and Industrial Innovations. In *Metrology and Standardization for Nanotechnology: Protocols  
565 and Industrial Innovations*, Mansfield, E.; Kaiser, D. L.; Fujita, D.; Van de Voorde, M., Eds.  
566 John Wiley & Sons: 2017; pp 411-464.
- 567 3. Harper, S.; Wohlleben, W.; Doa, M.; Nowack, B.; Clancy, S.; Canady, R.; Maynard, A.,  
568 Measuring nanomaterial release from carbon nanotube composites: review of the state of the  
569 science. *Journal of Physics: Conference Series* **2015**, *617*, (1), 012026.
- 570 4. Nowack, B.; David, R. M.; Fissan, H.; Morris, H.; Shatkin, J. A.; Stintz, M.; Zepp, R.;  
571 Brouwer, D., Potential release scenarios for carbon nanotubes used in composites. *Environ.Int.*  
572 **2013**, *59*, 1-11.
- 573 5. Sun, T.; Mitrano, D. M.; Bornhöft, N. A.; Scheringer, M.; Hungerbuehler, K.; Nowack,  
574 B., Envisioning nano release dynamics in a changing world: using dynamic probabilistic  
575 modelling to assess future environmental emissions of engineered nanoparticles. *Environmental  
576 Science & Technology* **2017**.
- 577 6. Garner, K. L.; Suh, S.; Keller, A. A., Assessing the Risk of Engineered Nanomaterials in  
578 the Environment: Development and Application of the nanoFate Model. *Environmental Science  
579 & Technology* **2017**, *51*, (10), 5541-5551.
- 580 7. Hund-Rinke, K.; Baun, A.; Cupi, D.; Fernandes, T. F.; Handy, R.; Kinross, J. H.; Navas,  
581 J. M.; Peijnenburg, W.; Schlich, K.; Shaw, B. J., Regulatory ecotoxicity testing of  
582 nanomaterials—proposed modifications of OECD test guidelines based on laboratory experience  
583 with silver and titanium dioxide nanoparticles. *Nanotoxicology* **2016**, *10*, (10), 1442-1447.
- 584 8. Duncan, T. V., Release of Engineered Nanomaterials from Polymer Nanocomposites: the  
585 Effects of Matrix Degradation. *ACS Applied Materials & Interfaces* **2014**, *7*, 20-39.
- 586 9. Duncan, T. V.; Pillai, K., Release of Engineered Nanomaterials from Polymer  
587 Nanocomposites: Diffusion, Dissolution, and Desorption. *ACS Applied Materials & Interfaces*  
588 **2014**, *7*, 1-19.
- 589 10. Mitrano, D. M.; Motellier, S.; Clavaguera, S.; Nowack, B., Review of nanomaterial aging  
590 and transformations through the life cycle of nano-enhanced products. *Environment  
591 International* **2015**, *77*, (0), 132-147.
- 592 11. Hüffer, T.; Praetorius, A.; Wagner, S.; Von Der Kammer, F.; Hofmann, T., Microplastic  
593 exposure assessment in aquatic environments: learning from similarities and differences to  
594 engineered nanoparticles. *Environmental Science & Technology* **2017**.
- 595 12. Koivisto, A. J.; Jensen, A. C. Ø.; Kling, K. I.; Nørgaard, A.; Brinch, A.; Christensen, F.;  
596 Jensen, K. A., Quantitative material releases from products and articles containing manufactured  
597 nanomaterials: Towards a release library. *NanoImpact* **2017**.
- 598 13. Nowack, B.; Boldrin, A.; Caballero, A.; Hansen, S. F.; Gottschalk, F.; Heggelund, L.;  
599 Hennig, M.; Mackevica, A.; Maes, H.; Navratilova, J., Meeting the Needs for Released  
600 Nanomaterials Required for Further Testing • The SUN Approach. *Environmental science &  
601 technology* **2016**, *50*, (6), 2747-2753.

- 602 14. Zhang, J.; Terrones, M.; Park, C. R.; Mukherjee, R.; Monthieux, M.; Koratkar, N.; Kim,  
603 Y. S.; Hurt, R.; Frackowiak, E.; Enoki, T.; Chen, Y.; Chen, Y.; Bianco, A., Carbon science in  
604 2016: Status, challenges and perspectives. *Carbon* **2016**, *98*, 708-732.
- 605 15. Ministère de l'Environnement, d. l. É. e. d. l. M., Éléments issus des déclarations des  
606 substances à l'état nanoparticulaire: Exercice 2015. **2015**.
- 607 16. Neubauer, N.; Scifo, L.; Navratilova, J.; Gondikas, A.; Mackevica, A.; Borschneck, D.;  
608 Chaurand, P.; Vidal, V.; Rose, J.; von der Kammer, F.; Wohlleben, W., Nanoscale Coloristic  
609 Pigments: Upper Limits on Releases from Pigmented Plastic during Environmental Aging, In  
610 Food Contact, and by Leaching. *Environmental Science & Technology* **2017**, *51*, (20), 11669-  
611 11680.
- 612 17. Hofmann, T.; Ma-Hock, L.; Strauss, V.; Treumann, S.; Rey Moreno, M.; Neubauer, N.;  
613 Wohlleben, W.; Gröters, S.; Wiench, K.; Veith, U.; Teubner, W.; van Ravenzwaay, B.;  
614 Landsiedel, R., Comparative short-term inhalation toxicity of five organic diketopyrrolopyrrole  
615 pigments and two inorganic iron-oxide-based pigments. *Inhalation Toxicology* **2016**, 1-17.
- 616 18. Ma-Hock, L.; Strauss, V.; Treumann, S.; Küttler, K.; Wohlleben, W.; Hofmann, T.;  
617 Gröters, S.; Wiench, K.; Ravenzwaay, B.; Landsiedel, R., Comparative inhalation toxicity of  
618 multi-wall carbon nanotubes, graphene, graphite nanoplatelets and low surface carbon black.  
619 *Part Fibre Toxicol* **2013**, *10*.
- 620 19. Wohlleben, W.; Brill, S.; Meier, M.; Mertler, M.; Cox, G.; Hirth, S.; von Vacano, B.;  
621 Strauss, V.; Treumann, S.; Wiench, K.; Ma-Hock, L.; Landsiedel, R., On the lifecycle of  
622 nanocomposites: comparing released fragments and their in-vivo hazards from three release  
623 mechanisms and four nanocomposites. *Small* **2011**, *7*, 2384 - 2395.
- 624 20. Wohlleben, W.; Kingston, C.; Carter, J.; Sahle-Demessie, E.; Vázquez-Campos, S.;  
625 Acrey, B.; Chen, C.-Y.; Walton, E.; Egenolf, H.; Müller, P.; Zepp, R., NanoRelease: Pilot  
626 interlaboratory comparison of a weathering protocol applied to resilient and labile polymers with  
627 and without embedded carbon nanotubes. *Carbon* **2017**, *113*, 346-360.
- 628 21. Wohlleben, W.; Neubauer, N., Quantitative rates of release from weathered  
629 nanocomposites are determined across 5 orders of magnitude by the matrix, modulated by the  
630 embedded nanomaterial. *NanoImpact* **2016**, *1*, 39-45.
- 631 22. Schlagenhaut, L.; Buerki-Thurnherr, T.; Kuo, Y.-Y.; Wichser, A.; Nüesch, F.; Wick, P.;  
632 Wang, J., Carbon Nanotubes Released from an Epoxy-Based Nanocomposite: Quantification and  
633 Particle Toxicity. *Environmental Science & Technology* **2015**, *49*, (17), 10616-10623.
- 634 23. OECD, Fish, Acute Toxicity Test (Annex 2 Composition of the recommended  
635 reconstituted water). . *Guidelines for Testing of Chemicals* **1992**, 203.
- 636 24. Bicho, R. C.; Santos, F. C.; Gonçalves, M. F.; Soares, A. M.; Amorim, M. J., Enchytraeid  
637 Reproduction TestPLUS: hatching, growth and full life cycle test—an optional multi-endpoint  
638 test with Enchytraeus crypticus. *Ecotoxicology* **2015**, *24*, (5), 1053-1063.
- 639 25. OECD, Guidance on sample preparation and dosimetry for the safety testing of  
640 manufactured nanomaterials. **2012**.
- 641 26. Bicho, R. C.; Ribeiro, T.; Rodrigues, N. P.; Scott-Fordsmand, J. J.; Amorim, M. J.,  
642 Effects of Ag nanomaterials (NM300K) and Ag salt (AgNO<sub>3</sub>) can be discriminated in a full life  
643 cycle long term test with Enchytraeus crypticus. *Journal of hazardous materials* **2016**, *318*, 608-  
644 614.
- 645 27. Petersen, E. J.; Diamond, S. A.; Kennedy, A. J.; Goss, G. G.; Ho, K.; Lead, J.; Hanna, S.  
646 K.; Hartmann, N. B.; Hund-Rinke, K.; Mader, B.; Manier, N.; Pandard, P.; Salinas, E. R.; Sayre,  
647 P., Adapting OECD Aquatic Toxicity Tests for Use with Manufactured Nanomaterials: Key

648 Issues and Consensus Recommendations. *Environmental Science & Technology* **2015**, *49*, (16),  
649 9532-9547.

650 28. Lin, S.; Zhao, Y.; Ji, Z.; Ear, J.; Chang, C. H.; Zhang, H.; Low-Kam, C.; Yamada, K.;  
651 Meng, H.; Wang, X., Zebrafish high-throughput screening to study the impact of dissolvable  
652 metal oxide nanoparticles on the hatching enzyme, ZHE1. *Small* **2013**, *9*, (9-10), 1776-1785.

653 29. Lin, S.; Zhao, Y.; Xia, T.; Meng, H.; Ji, Z.; Liu, R.; George, S.; Xiong, S.; Wang, X.;  
654 Zhang, H., High content screening in zebrafish speeds up hazard ranking of transition metal  
655 oxide nanoparticles. *ACS nano* **2011**, *5*, (9), 7284-7295.

656 30. Lin, S.; Wang, X.; Ji, Z.; Chang, C. H.; Dong, Y.; Meng, H.; Liao, Y.-P.; Wang, M.;  
657 Song, T.-B.; Kohan, S., Aspect ratio plays a role in the hazard potential of CeO<sub>2</sub> nanoparticles in  
658 mouse lung and zebrafish gastrointestinal tract. *ACS nano* **2014**, *8*, (5), 4450-4464.

659 31. Lee, L. E.; Clemons, J. H.; Bechtel, D. G.; Caldwell, S. J.; Han, K.-B.; Pasitschniak-Arts,  
660 M.; Mosser, D. D.; Bols, N. C., Development and characterization of a rainbow trout liver cell  
661 line expressing cytochrome P450-dependent monooxygenase activity. *Cell biology and*  
662 *toxicology* **1993**, *9*, (3), 279-294.

663 32. Faisal, M.; Ahne, W., A cell line (CLC) of adherent peripheral blood mononuclear  
664 leucocytes of normal common carp *Cyprinus carpio*. *Developmental & Comparative*  
665 *Immunology* **1990**, *14*, (2), 255-260.

666 33. Weyts, F.; Rombout, J.; Flik, G.; Verburg-Van Kemenade, B., A common carp (*Cyprinus*  
667 *carpio*L.) leucocyte cell line shares morphological and functional characteristics with  
668 macrophages. *Fish & Shellfish Immunology* **1997**, *7*, (2), 123-133.

669 34. Lammel, T.; Boisseaux, P.; Fernández-Cruz, M.-L.; Navas, J. M., Internalization and  
670 cytotoxicity of graphene oxide and carboxyl graphene nanoplatelets in the human hepatocellular  
671 carcinoma cell line Hep G2. *Particle and fibre toxicology* **2013**, *10*, (1), 27.

672 35. Lammel, T.; Navas, J. M., Graphene nanoplatelets spontaneously translocate into the  
673 cytosol and physically interact with cellular organelles in the fish cell line PLHC-1. *Aquatic*  
674 *toxicology* **2014**, *150*, 55-65.

675 36. Schlagenhauf, L.; Kuo, Y.-Y.; Bahk, Y. K.; Nüesch, F.; Wang, J., Decomposition and  
676 particle release of a carbon nanotube/epoxy nanocomposite at elevated temperatures. *Journal of*  
677 *Nanoparticle Research* **2015**, *17*, (11), 1-11.

678 37. Singh, D.; Schiffman, L. A.; Watson-Wright, C.; Sotiriou, G. A.; Oyanedel-Craver, V.;  
679 Wohlleben, W.; Demokritou, P., Nanofiller presence enhances polycyclic aromatic hydrocarbon  
680 (PAH) profile on nanoparticles released during thermal decomposition of nano-enabled  
681 thermoplastics: Potential Environmental Health Implications. *Environmental Science &*  
682 *Technology* **2017**.

683 38. Sotiriou, G. A.; Singh, D.; Zhang, F.; Chalbot, M.-C. G.; Spielman-Sun, E.; Hoering, L.;  
684 Kavouras, I. G.; Lowry, G. V.; Wohlleben, W.; Demokritou, P., Thermal decomposition of nano-  
685 enabled thermoplastics: Possible environmental health and safety implications. *Journal of*  
686 *Hazardous Materials* **2016**, *305*, 87-95.

687 39. Hirth, S.; Cena, L. G.; Cox, G.; Tomovic, Z.; Peters, T. M.; Wohlleben, W., Scenarios  
688 and methods that induce protruding or released CNTs after degradation of composite materials.  
689 *J.Nanoparticle Res.* **2013**, *15*, 1504.

690 40. Kang, J.; Erdely, A.; Afshari, A.; Casuccio, G.; Bunker, K.; Lersch, T.; Dahm, M.;  
691 Farcas, D.; Cena, L., Generation and characterization of aerosols released from sanding  
692 composite nanomaterials containing carbon nanotubes. *NanoImpact* **2016**.

- 693 41. Watson, C.; DeLoid, G.; Pal, A.; Demokritou, P., Buoyant Nanoparticles: Implications  
694 for Nano-Biointeractions in Cellular Studies. *Small* **2016**.
- 695 42. Watson-Wright, C.; Singh, D.; Demokritou, P., Toxicological implications of released  
696 particulate matter during thermal decomposition of nano-enabled thermoplastics. *NanoImpact*  
697 **2016**.
- 698 43. Santos, F. C.; Gomes, S. I.; Scott-Fordsmand, J. J.; Amorim, M. J.; Freitas, V., Hazard  
699 assessment of Nickel nanoparticles in soil - the use of a full life cycle test with *Enchytraeus*  
700 *crypticus*. *Environmental Toxicology and Chemistry* **2017**.
- 701 44. Scott-Fordsmand, J. J.; Navas, J. M.; Hund-Rinke, K.; Nowack, B.; Amorim, M. J. B.,  
702 Nanomaterials to microplastics: Swings and roundabouts. *Nano Today* **2017**.
- 703 45. Störmer, A.; Bott, J.; Kemmer, D.; Franz, R., Critical review of the migration potential of  
704 nanoparticles in food contact plastics. *Trends in Food Science & Technology* **2017**, *63*, 39-50.
- 705 46. Sussarellu, R.; Suquet, M.; Thomas, Y.; Lambert, C.; Fabioux, C.; Pernet, M. E. J.; Le  
706 Goïc, N.; Quillien, V.; Mingant, C.; Epelboin, Y.; Corporeau, C.; Guyomarch, J.; Robbins, J.;  
707 Paul-Pont, I.; Soudant, P.; Huvet, A., Oyster reproduction is affected by exposure to polystyrene  
708 microplastics. *Proceedings of the National Academy of Sciences* **2016**, *113*, (9), 2430-2435.
- 709 47. Avio, C. G.; Gorbi, S.; Milan, M.; Benedetti, M.; Fattorini, D.; d'Errico, G.; Pauletto, M.;  
710 Bargelloni, L.; Regoli, F., Pollutants bioavailability and toxicological risk from microplastics to  
711 marine mussels. *Environmental Pollution* **2015**, *198*, (Supplement C), 211-222.
- 712 48. Ging, J.; Tejerina-Anton, R.; Ramakrishnan, G.; Nielsen, M.; Murphy, K.; Gorham, J.  
713 M.; Nguyen, T.; Orlov, A., Development of a conceptual framework for evaluation of  
714 nanomaterials release from nanocomposites: Environmental and toxicological implications.  
715 *Science of The Total Environment* **2014**, *473*, 9-19.
- 716 49. Gomez, V.; Levin, M.; Saber, A. T.; Irusta, S.; Dal Maso, M.; Hanoi, R.; Santamaria, J.;  
717 Jensen, K. A.; Wallin, H.; Koponen, I. K., Comparison of Dust Release from Epoxy and Paint  
718 Nanocomposites and Conventional Products during Sanding and Sawing. *Annals of*  
719 *Occupational Hygiene* **2014**.
- 720 50. Saber, A.; Koponen, I.; Jensen, K.; Jacobsen, N.; Mikkelsen, L.; Moller, P.; Loft, S.;  
721 Vogel, U.; Wallin, H., Inflammatory and genotoxic effects of sanding dust generated from  
722 nanoparticle-containing paints and lacquers. *Nanotoxicology* **2012**, *6*, 776 - 788.
- 723 51. Saber, A.; Jacobsen, N.; Mortensen, A.; Szarek, J.; Jackson, P.; Madsen, A.; Jensen, K.;  
724 Koponen, I.; Brunborg, G.; Gutzkow, K.; Vogel, U.; Wallin, H., Nanotitanium dioxide toxicity in  
725 mouse lung is reduced in sanding dust from paint. *Part.Fibre Toxicol.* **2012**, *9*, (1), 4.
- 726 52. Wohlleben, W.; Meier, M. W.; Vogel, S.; Landsiedel, R.; Cox, G.; Hirth, S.; Tomović,  
727 Ž., Elastic CNT–polyurethane nanocomposite: synthesis, performance and assessment of  
728 fragments released during use. *Nanoscale* **2013**, *5*, (1), 369-380.
- 729 53. Saber, A. T.; Mortensen, A.; Szarek, J.; Koponen, I. K.; Levin, M.; Jacobsen, N. R.;  
730 Pozzebon, M. E.; Mucelli, S. P.; Rickerby, D. G.; Kling, K.; Atluri, R.; Madsen, A. M.; Jackson,  
731 P.; Kyjovska, Z. O.; Vogel, U.; Jensen, K. A.; Wallin, H., Epoxy composite dusts with and  
732 without carbon nanotubes cause similar pulmonary responses, but differences in liver histology  
733 in mice following pulmonary deposition. *Particle and Fibre Toxicology* **2016**, *13*, (1), 1-20.
- 734 54. Kaiser, J. P.; Roesslein, M.; Diener, L.; Wick, P., Human Health Risk of Ingested  
735 Nanoparticles That Are Added as Multifunctional Agents to Paints: an In Vitro Study. *PLOS*  
736 *ONE* **2013**, *8*, (12), e83215.

- 737 55. Semenzin, E.; Hristozov, D.; Caballero, A.; Scott-Fordsmand, J.; Gottschalk, F.; Basei,  
738 G. P.; Subramanian, V.; Zabeo, A.; Nowack, B.; Marcomini, A., Ecological risk along the life-  
739 cycle of nano-enabled products. *in preparation* **2017**.
- 740 56. V. Pillai, K.; Gray, P. J.; Tien, C.-C.; Bleher, R.; Sung, L.-P.; V. Duncan, T.,  
741 Environmental release of core-shell semiconductor nanocrystals from free-standing polymer  
742 nanocomposite films. *Environmental Science: Nano* **2016**, 3, (3), 657-669.
- 743 57. Mitrano, D. M.; Mehrabi, K.; Arroyo, Y.; Nowack, B., Mobility of metallic  
744 (nano)particles in leachates from landfills containing waste incineration residues. *Environmental*  
745 *Science: Nano* **2016**.
- 746 58. Mitrano, D. M.; Limpiteprakan, P.; Babel, S.; Nowack, B., Durability of nano-enhanced  
747 textiles through the life cycle: releases from landfilling after washing. *Environmental Science:*  
748 *Nano* **2016**, 3, (2), 375-387.
- 749 59. Wohlleben, W.; Meyer, J.; Muller, J.; Muller, P.; Vilsmeier, K.; Stahlmecke, B.;  
750 Kuhlbusch, T. A. J., Release from nanomaterials during their use phase: combined mechanical  
751 and chemical stresses applied to simple and multi-filler nanocomposites mimicking wear of  
752 nano-reinforced tires. *Environmental Science: Nano* **2016**, 3, 1036-1051.
- 753 60. Lassen, C.; Hansen, S. F.; Magnusson, K.; Norèn, F.; Bloch Hartmann, N. I.; Jensen, P.  
754 R.; Nielsen, T. G.; Brinch, A. *Microplastics - Occurrence, effects and sources of releases to the*  
755 *environment in Denmark*; 2015; pp 1-205.
- 756 61. Taylor, M. L.; Gwinnett, C.; Robinson, L. F.; Woodall, L. C., Plastic microfibre ingestion  
757 by deep-sea organisms. *Scientific Reports* **2016**, 6, 33997.
- 758 62. Deng, Y.; Zhang, Y.; Lemos, B.; Ren, H., Tissue accumulation of microplastics in mice  
759 and biomarker responses suggest widespread health risks of exposure. *Scientific Reports* **2017**, 7,  
760 46687.
- 761