Short communication

A new biogenerated Rh-based catalyst for aqueous biphasic hydroformylation

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A new bio-generated rhodium based system embedded in a peculiar polysaccharide matrix (\textit{Rh-EPS}), was obtained and purified from cultures of bacterial cells of \textit{Klebsiella oxytoca} DSM 29614. The product was analyzed with different techniques to obtain information on its structure–property correlation. In order to determine its catalytic activity and selectivity in the aqueous biphasic hydroformylation some olefins were chosen as model substrates, obtaining fine-good results.

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1. Introduction

Hydroformylation is one of the most important industrial reactions catalyzed by soluble metal complexes \cite{1}. The most efficient catalysts in terms of both activity and selectivity are rhodium complexes, capable to operate under mild conditions \cite{1}. The major drawback of this homogeneous process is the separation of the expensive catalyst from the product mixture that requires an energy intensive process such as distillation. For this reason and for environmental aspects of the chemical production, liquid–liquid two-phase systems have been developed over the last years, with the catalyst confined in one phase and the product in the other one \cite{2,3}. In particular, the use of environmentally more benign solvents as water has been developed \cite{2,3} and the hydroformylation process represents one of the most striking examples of this catalytic methodology \cite{1–4}. The use of natural compounds, such as aminocids, peptides, proteins and sugars as ligands to maintain metallic species soluble and active in aqueous biphasic systems was deeply investigated \cite{5–15}. Since bacterial cells may often synthesize and secrete polysaccharides \cite{16}, recently we envisaged a new way of using bacteria to produce carbohydrates able to bind metals during their growing in the presence of suitable metal salts. In particular \textit{Klebsiella oxytoca} DSM 29614 (ex strain BAS-10) strain, isolated from the acid mine drainage of pyrite mines and able to produce a peculiar exopolysaccharide (\textit{EPS}), constituted by a branched heptameric repeating structure \cite{17}, was investigated with success by some of us to generate some metal-polysaccharide catalysts \cite{18,19}. In this study, a new bio-generated Rh based catalyst (\textit{Rh-EPS}) was produced in order to determine its activity and selectivity in the aqueous biphasic hydroformylation of some olefins chosen as model substrates.

2. Experimental section

2.1. Materials and instrumentation

\(\text{NaHCO}_3, \text{NH}_4\text{Cl}, \text{MgSO}_4 \cdot 7\text{H}_2\text{O}, \text{NaH}_2\text{PO}_4, \text{KCl}, \text{Na-citrate}, \text{styrene, 1-octene, and toluene were Aldrich products. 2-allyl-5-ethylthiophene (IX) and 1-[(5-ethylthiophen-2-yl)propene (XII) were prepared with 60% and 92% yields, respectively, using a Pd-catalyzed cross-coupling reaction of the Grignard reagent obtained from Chimet S.p.A. (Italy). GC analyses were carried out on an Agilent 6850A gaschromatograph (HP1 column 30 m × 0.32 mm × 0.25 \mu m). Atomic absorption measurements were carried out by using a Perkin Elmer AAANALYST 100 instrument. TEM analyses were performed on a JEOL JEM 100b microscope. IR spectra (KBr pellets) were recorded on an FTIR Nicolet Magna 750 instrument.}

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2.2. Biogenerated catalyst preparation

2.2.1. Anaerobic cultivation

The K. oxytoca DSM 29614 was retrieved from cryovials kept at −80 °C in 25% glycerol in Nutrient broth (Difco). The production of Rh-EPS was made in NAC medium containing per liter: 2.5 g NaHCO₃, 1.5 g NH₄Cl, 1.5 g MgSO₄·7H₂O, 0.6 g NaH₂PO₄, 0.1 g KCl and 50 mM (14.7 g·L⁻¹) Na-citrate, and finally it was buffered at pH 7.6 with NaOH. The NAC medium prepared in 1 l Pyrex bottle was cooled down by N₂ fluxing. After 1 day of stationary growth phase (2 days) the culture was amended with 50 mg of total rhodium as RhCl₃. The medium was kept under anaerobic conditions by N₂ fluxing. At the end rhodium, bound to the exopolysaccharide, precipitated from the culture to the bottom bottle as Rh-EPS.

2.3. Rh-EPS purification

Bacterial cell residues were eliminated by centrifugation at 4000 g per 25’ and then the supernatant was treated with 800 ml of cooled ethyl alcohol (95%) to precipitate all Rh-EPS. Salt residues were removed by washing Rh-EPS three times with distilled water. The colloidal material was dried out under vacuum to obtain Rh-EPS, as a solid dry material, and was stored at 4 °C.

2.4. Biogenerated catalyst characterization

2.4.1. Content of Rh in Rh-EPS

A Rh-EPS sample was analyzed by atomic absorption after treatment with aqua regia to determine the total Rh concentration: the obtained value was 0.8%.

2.4.2. Transmission electron microscope observations

To observe the electron dense rhodium in EPS, several specimens were prepared for transmission electron microscopy (TEM). An amount of 10 mg of dried catalyst was suspended in 5 ml distilled water and, after 10 min in ultrasonic bath, 10 μl of the suspension was mounted on a platinum grid which was previously coated with a Formvar bio film. The liquid was evaporated at room temperature and the layered specimen on the grid was observed by transmission electron microscopy operating under standard conditions.

2.4.3. X-ray photoelectron spectroscopy

XPS was used as the main tool to assess the surface chemical composition of the samples before and after the catalytic process. Analyses were performed on a Perkin-Elmer Φ 5600-ci spectrometer using non-monochromatized Al Kα radiation (1486.6 eV). The spectrometer was calibrated by assuming the binding energy (BE) of the Au 4f7/2 line at 83.9 eV with respect to the Fermi level. Survey scans were obtained in the 0–1300 eV range. Detailed scans were recorded for the Rh3d, N1s, O1s, P2p, Mg2s, Na1s and C1s regions. The BEs values were corrected for charging effects by assigning to the C1s peak associated with adventitious hydrocarbons a value of 284.8 eV [22]. Samples were mounted as finely grounded powders on steel holders and introduced directly in the fast-entry lock system of the XPS analytical chamber. No sizeable sign of sample degradation was observed under X-ray irradiation.

2.4.4. Powder X-ray diffraction

PXRD was used to investigate the crystalline phases before and after the catalytic process. PXRD measurements were carried out by means of a Bruker D8 Advance diffractometer equipped with a Göbel mirror and a Cu-Kα source (40 kV, 40 mA). PXRD patterns of struvite and bohierite were simulated by means of Mercury 3.5.1 software with a full width at half maximum (FWHM) of 0.3° and 0.5° (in 2θ), respectively.

2.5. Aqueous biphasic hydroformylation experiments

All the reactions were carried out following a procedure similar to that below described for the Rh-EPS catalyzed hydroformylation of styrene (I). Experimental details are reported in Tables 1–3.

2.5.1. Aqueous biphasic hydroformylation of styrene (I)

In a Schlenk tube, 3.4 mg (0.26 mmol) of Rh-EPS were stirred under nitrogen in 2 ml of distilled water for about 10’. A solution of 27.0 mg (0.26 mmol) of freshly distilled styrene (I) in 2 ml of toluene was then added to the aqueous phase. The Schlenk tube was then transferred into a 150 ml stainless steel autoclave under nitrogen, pressurized with 6 MPa of syngas (CO/H₂ = 1) and stirred for 18 h at 60 °C (Table 1). The reactor was then cooled to room temperature and the residual gases released. Diethyl ether was added and the organic phase was separated, dried on Na₂SO₄ and analyzed by GC and GC–MS. The catalytic aqueous phase was recycled for further experiments after addition of fresh styrene (I).

3. Results and discussion

The strain K. oxytoca DSM 29614 produces an exopolysaccharide (EPS) with the following structure: 4 rhamnose (Rha), 2 glucuronic acids (GlcA) and 1 galactose (Gal) bound by glycosidic bonds as following [17]:

\[2\alpha\text{-}\text{Rha}-(1\rightarrow 3)\beta\text{-}\text{Gal}-(1\rightarrow 2)\alpha\text{-}\text{Rha}-(1\rightarrow 4)\beta\text{-}\text{GlcA}-(1\rightarrow 3)\alpha\text{-}\text{Rha}-(1\rightarrow 4)\text{-}\text{EPS} \]

\[\beta\text{-}\text{GlcA} \]

which has neither glucose nor fucose, which are in general present in common colanic acid [23]. The latter in Enterobacteriaceae is a polyanionic heteropolysaccharide containing a repeat unit with β-glucose, L-fucose, D-galactose, and β-glucuronate sugars. This anomalous colanic acid analogue secreted by K. oxytoca is likely the result of adaptation to high concentrations of heavy metals, typical of acid drainage of pyrite mines. EPS binds iron and other metals during the bacterial growth, as defense mechanism to metal poisoning [23]. The metal is sequestered by the exopolymeric structure, which prevents the metal to

<table>
<thead>
<tr>
<th>Entry</th>
<th>Substrate</th>
<th>Conv. (%)</th>
<th>VI yield (%)</th>
<th>VII yield (%)</th>
<th>VIII yield (%)</th>
<th>V yield (%)</th>
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</tr>
<tr>
<td>4</td>
<td>IV</td>
<td>0</td>
<td>-</td>
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<td>V</td>
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<td>14.9</td>
<td>5.7</td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>V</td>
<td>18.6</td>
<td>nd</td>
<td>13.9</td>
<td>4.7</td>
<td></td>
</tr>
</tbody>
</table>

Reaction conditions: substrate = 0.65 mmol; substrate/Rh (molar ratio) = 1000/1; p(CO) = p(H₂) = 3 MPa; T = 60 °C; t = 22 h; H₂O = 2 ml; Toluene = 2 ml. α Experiment carried out by using the catalytic phase recovered from the previous run.

b H₂O = 0 ml; Toluene = 4 ml. nd = not detected in the reaction mixture.

Table 2

Aqueous biphasic hydroformylation of 1-ocetene (IV) and 2-ocetene (V) catalyzed by Rh-EPS.

<table>
<thead>
<tr>
<th>Entry</th>
<th>Substrate</th>
<th>Conv. (%)</th>
<th>VI yield (%)</th>
<th>VII yield (%)</th>
<th>VIII yield (%)</th>
<th>V yield (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>IV</td>
<td>99.0</td>
<td>54.2</td>
<td>36.5</td>
<td>1.4</td>
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<td>3</td>
<td>IV</td>
<td>95.5</td>
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<tr>
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<td>IV</td>
<td>0</td>
<td>-</td>
<td>-</td>
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<td>-</td>
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<tr>
<td>5</td>
<td>V</td>
<td>20.6</td>
<td>nd</td>
<td>14.9</td>
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<td>6</td>
<td>V</td>
<td>18.6</td>
<td>nd</td>
<td>13.9</td>
<td>4.7</td>
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</table>

Reaction conditions: substrate = 0.65 mmol; substrate/Rh (molar ratio) = 1000/1; p(CO) = p(H₂) = 3 MPa; T = 60 °C; t = 22 h; H₂O = 2 ml; Toluene = 2 ml. α Experiment carried out by using the catalytic phase recovered from the previous run.

b H₂O = 0 ml; Toluene = 4 ml. nd = not detected in the reaction mixture.
come into direct contact with cell membranes and even less with the cytoplasm. To support the hypothesis that also in this case rhodium is embedded in the polysugar moiety we compared the IR spectra of Rh-EPS and EPS (Fig. 1); they show different patterns, and the most evident difference is the peak at about 1000 cm\(^{-1}\) in Rh-EPS due to the asymmetric stretching of a P–O–C group of an organic phosphate\[24\].

The addition of rhodium probably causes the formation of organic phosphates because of an intense phosphatase activity (data not shown) which is likely triggered by Rh\(^{3+}\) toxicity. The precipitation of metals by phosphates was found also in \textit{Citrobacter} sp \[25,26\]. So the secretion of exopolysaccharide and phosphatase activity explains the high resistance of DSM 29614 to heavy metal cations. XPS analysis on Rh-EPS confirmed the presence of phosphorous along with carbon, nitrogen, alkaline and alkaline-earth cations (Na\(^{+}\), Mg\(^{2+}\)). The P2p line is centered at BE values (ca. 133 eV) typical for phosphate species \[27\] and the N1s region at 400.3 eV is associated with ammonium salts. At variance, unfortunately, rhodium is not revealed likely due to its low concentration.

To confirm the presence of inorganic phosphates in Rh-EPS and to investigate the crystalline phases before and after the catalytic process PXRD characterization was performed. The Rh-EPS PXRD pattern before (a) and after (b) the hydroformylation experiments is reported in Supplementary material. The major crystalline phase in the as prepared catalyst is ascribed to \textit{struvite}, i.e. MgNH\(_4\)PO\(_4\) · (H\(_2\)O)\(_6\), JCPDS card no. 71-2089 (a). The \textit{struvite} simulated PXRD pattern (FWHM = 0.3\(^\circ\)) well reproduces the major peaks. Some peaks, marked with asterisks, are not ascribed to MgNH\(_4\)PO\(_4\) · (H\(_2\)O)\(_6\) but could be related to the presence of a mixture of minor crystalline phases of sodium phosphate and sodium hydrogen phosphate hydrate or to the presence of organic phosphates. After the catalytic process Rh-EPS PXRD pattern changes considerably. The two most intense peaks centered at 2\(\theta\) = 11.1\(^\circ\) and 12.6\(^\circ\) (b) are indexed as the (021) and (040) reflections of the \textit{bobieriite}, i.e. Mg\(_3\)(PO\(_4\))\(_2\) · (H\(_2\)O)\(_6\), monoclinic structure (JCPDS card no. 16-0330). Also in this case, the \textit{bobieriite} simulated PXRD pattern (FWHM = 0.5\(^\circ\)) reproduces very well the experimental data. Hence, after the hydroformylation the major crystalline phase changes from an ammonium magnesium phosphate hydrate to a magnesium phosphate hydrate. PXRD analysis again do not show any signal ascribable to rhodium nanoparticles. So this type of analysis was not conclusive to define the interaction of rhodium species with EPS.

More diagnostic were TEM micrographs that show the formation of rhodium nanoparticles embedded in an exopolysaccharidic matrix (Rh-EPS NPs) (Fig. 2). In the inset a magnification of a nanoparticle shows an irregular shape probably due to the presence of nanocrystals of phosphates with elongated dendritic morphology \[28\] associated with rhodium and exopolysaccharide.

### Table 3

Aqueous biphasic hydroformylation of 2-allyl-5-ethylthiophene (IX) catalyzed by Rh-EPS.

<table>
<thead>
<tr>
<th>Entry</th>
<th>Conv. (%)</th>
<th>X yield (%)</th>
<th>XI yield (%)</th>
<th>XII yield (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>96.0</td>
<td>47.0</td>
<td>45.5</td>
<td>3.5</td>
</tr>
<tr>
<td>2(^a)</td>
<td>23.4</td>
<td>12.0</td>
<td>11.4</td>
<td>nd</td>
</tr>
<tr>
<td>3(^a)</td>
<td>8.2</td>
<td>4.5</td>
<td>3.7</td>
<td>nd</td>
</tr>
</tbody>
</table>

**Reaction conditions:** substrate = 0.65 mmol; substrate/Rh (molar ratio) = 1000/1; p(CO) = p(H\(_2\)) = 3.5 MPa; T = 60 °C; t = 22 h; H\(_2\)O = 2 ml; toluene = 2 ml.

\(^{a}\) Experiment carried out by using the catalytic phase recovered from the previous run.

nd = not detected in the reaction mixture.

3.1. Catalytic properties of Rh-EPS

A first set of o xo-experiments was performed on styrene (I) as this aromatic olefin is a widespread studied model substrate for functionalized olefins \[1\] (Scheme 1). The hydroformylation reactions were carried out in the biphasic medium water/toluene, in the presence of the catalytic system Rh-EPS at 6 MPa of syngas (CO/H\(_2\) = 1) and 60 °C for 18 h with a substrate to rhodium molar ratio = 1000. The catalytic aqueous phase was recycled in three consecutive experiments and the data obtained are reported in Table 1. The catalytic system was very active, showing complete conversion and chemoselectivity: neither ethylbenzene nor alcohols were formed: also the regioselectivity towards the branched aldehyde, 2-phenylpropanal (II), was very high (93–94%). The...
catalyst activity remained practically unchanged in the first recycle but, unfortunately, a strong decrease was observed starting from the second recycle. Because of the low amount, the decreased activity of the catalyst can be tentatively ascribed both to a mechanical loss and/or to an accidental poisoning due to the presence of oxygen during the phase separation. In all cases, however, chemo- and regioselectivity did not show any significant difference. Noteworthy, when the reaction was carried out at 60 °C, 3 MPa of CO and 3 MPa of H2 for 22 h, conversion was very high (98%) and, as expected, n-nonanal (VI) was the main product. Besides the two major aldehydes VI and VII, also a very small amount of the isomeric aldehyde VIII was formed, due to the isomerisation of the carbon–carboxy double bond: as a matter of fact about 7% of 2-octene (V) was detected in the reaction mixture (Entry 1, Table 2). The catalytic phase was reused in two consecutive recycling experiments and its activity was maintained practically unchanged. Noteworthy, analogously to what was observed in the hydrogenation process catalyzed by Rh-EPS [19], also Rh-EPS, though not soluble in water but only suspended in it, is active exclusively in the presence of water. As a matter of fact, when toluene was used as the only reaction medium, no reaction occurred (Entry 4, Table 2). We hypothesize that water swelling of the polymer structure makes rhodium more available for the catalysis process.

Despite its complex structure this catalytic system was shown to be able to hydroformylate also internal double bonds, even if at a low reaction rate; indeed, 2-octene (V) was subjected to the oxo-process at the above reaction conditions and aldehydes VII and VIII were the sole reaction products (Entry 5, Table 2). The aqueous catalytic phase, when used in a recycling experiment, maintained its activity almost unchanged (Entry 6, Table 2).

Due to our interest in the synthesis of fine chemicals, in particular of fragrances [8,9,19], we finally tested the activity of Rh-EPS in the hydroformylation of 2-allyl-5-ethylthiophene (IX) (Scheme 3, Table 3), being known that some thiophene aldehydes, as compound X for instance, structurally similar to the fragrance Lioral® [32], have a relevant commercial interest.

The reaction was carried out in the biphasic water/toluene system at 60 °C and 7 MPa of syngas (CO/H2 = 1) for 24 h. In these conditions, conversion was very high (96%) and, surprisingly, the two aldehydes X and XI were obtained in almost equimolar amounts; moreover, also a small amount of the isomerized olefin XII was detected in the mixture. To our knowledge, olefin IX has never been hydroformylated and the formation of the branched aldehyde X, sterically unfavoured, as the prevailing product, is rather intriguing. We could infer this result to the isomerizing capability of our catalytic system, demonstrated also by the presence of the olefin XII in the reaction mixture. However, when

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**Scheme 1.** Hydroformylation of styrene (I).

**Scheme 2.** Hydroformylation of 1-octene (IV) and 2-octene (V).

**Fig. 2.** TEM micrograph of EPS containing Rhodium nanoparticles (electron dense particles), bar = 500 nm. (In set) one elongated particle with 150 × 90 nm size.
and 2-(1-phenylvinyl)pyridine. Moreover, the internal olefins rendered substrates as 1,1-diphenylethene, 1,3-diisopropenylbenzene and 2-(1-phenylvinyl)pyridine. We tried to hydroformylate olefins as Rh-EPS, rendering more difficult interaction with a complex catalytic system as Rh-EPS. Therefore, we can ascribe the high amount of the branched aldehyde X to the great affinity of the “soft” metal rhodium for the sulfur atom [33]: the coordination of S to Rh could help the interaction of the catalytically active metal with the carbon atom in β-position to the thiophenyl ring. The aqueous phase containing Rh-EPS was reused for two consecutive experiments but, unfortunately, the activity of the catalytic system strongly decreased since the first recycling reaction (Entry 2, Table 3). Up to now we have no clear explanation of this phenomenon but we can hypothesize a sulfur poisoning of the catalyst via a competitive absorption onto rhodium metal active sites[34,35]; in particular the thiophene ring may affect the weak coordination of the metal to the oxygen groups present in the sugar, so determining a change in the structure of the catalyst that results negative for the activity.

4. Conclusions

Our strategy to generate metal binding exopolysaccharides in the direction of developing new catalysts able to work under biphasic water-organic solvent conditions was applied with success to prepare a new rhodium species which catalyzed the hydroformylation reaction of some model olefins. Peculiar stereo- and regio-selectivity with aliphatic olefins and with olefins containing a heteroaryl moiety in beta position was observed while no activity was found with some sterically hindered substrates. In some cases a recycling ability of the catalyst was found.

Supplementary data to this article can be found online at http://dx.doi.org/10.1016/j.catcom.2015.08.001.

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